RELATIONSHIPS AMONG SPECIFIC GROWTH, SECRETED ACID PHOSPHATASE, INTRACELLULAR AND TOTAL INORGANIC PHOSPHORUS CONCENTRATION IN CHLORELLA FUSCA CULTURED UNDER VARIOUS LEVELS OF Na2HPO4

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Abstract
The relationships among specific growth rates, intracellular inorganic phosphate (Pi) and total phosphorus concentrations and specific activities of acid phosphatase (acPase) were studied on the green unicellular alga Chlorella fusca at 1.0, 15.0 and 30.0 μM Na2HPO4. The specific growth, the intracellular Pi and total phosphorus concentration increased with increasing Na2HPO4 levels. The acid phosphatase activity was inversely correlated with the specific growth rate, the total P concentrations and the intracellular inorganic concentrations. The P-deficiency induction of acPase activity is related to a decrease in P-availability in Chlorella fusca.

Key words: Acid phosphatase - Growth - Phosphorus

Introduction
Phosphorus is an important structural constituent of many biomolecules such as proteins, phospholipids and nucleic acids; therefore, its regulation is important for plant growth and development (Duff et al., 1994). Phosphorus also required for energy transfer and regulation of metabolism constituting from 0.3 to 0.5 % of plant dry weight (Cashikar et al., 1997). Under conditions of phosphate limitation, the growth and development of plants are particularly dependent upon the availability of phosphates (Dracup et al., 1984).

Among the various dissolved forms of phosphorus, orthophosphates (Pi) is known to be the most biologically available and readily usable form by phytoplankton (Cembella et al., 1984 and Bostrom et al., 1988).

Exposure of algae to phosphorus deficient waters, results in disorders in the metabolism such as decrease in growth, photosynthesis and respiration rate (Davis, 1988; Theodorou et al., 1991 and Garcia-Sanchez et al., 1996).

Phosphatases are a group of enzymes that catalyze the hydrolysis of a large number of phosphomonoesters. It is generally thought that the hydrolysis process that is mediated by phosphatase is related to the availability of Pi. In the blue green alga Anabaena flos-aquae, Healey (1973) reported that the removal of...
Pi from the medium was accompanied by increasing alkaline phosphatase activity. In the flagellate alga *Euglena gracilis*, the activity of acid phosphatase, also increased under P-deficient conditions (Price, 1962 and Blum, 1965). The induction of phosphatase activity is therefore considered a typical response in algae exposed to P-deficiency.

Orhanovic and Pavella-Vrancic (2000) emphasized that concentration of inorganic phosphorus in culture media controls the biosynthesis of the phosphatases and that the enzyme activity can often be used as an indicator of the Pi requirement in the planktonic community.

Despite the fact that phosphorus is an important and often limiting nutrient there is little knowledge on the nature and regulation of phosphatases in freshwater algae. Alkaline phosphatases were more pronounced than acid phosphatases in the literature cited.

The relationship between the acid phosphatase activity from algal species and the concentration of the available inorganic phosphate has been detected in several reports; Price (1962) and Blum (1965) on the flagellate alga *Euglena gracilis*, Sommer and Blum (1965); Moller et al., (1975) on microalgae. Weich and Groneli (1989) on *Ulva lactuca*; Quisel et al., (1996) on *Chlamydomonas reinhardtii*; Lee (2000 and 2004) on the red macroalga *Gracillaria coronopifolia*.

The present study deals with determination of the relationship among specific growth rate, intracellular inorganic phosphorus and total phosphorus concentrations as well as the specific activity of extracellular phosphatases isolated from the freshwater green alga *Chlorella fusca*.

**Materials and methods**

1- The tested organism:

*Chlorella fusca* was obtained from the culture collection of Gottingen University, Germany. It was maintained in the laboratory on a selective solid medium (Grimme and Boardman, 1972) for subsequent use. All the experiments performed for this alga were done in aerated liquid cultures. Cells were harvested by centrifugation at 3000 rpm, the pellets were left to dry on plastic foil and then divided each into two parts; the first one was used for determination of growth parameters and phosphatase activity while the other part was stored in the deep freezer for subsequent isolation and biochemical characterization of phosphatase enzyme.

2- Isolation of fractions enriched in secreted acid phosphatase:

Freezed tissues were used for preparing fractions of the secreted acid phosphatases. About 0.6 g fresh weight of the alga were suspended in 5 ml Chu 10 medium, shaken gently and pelleted by centrifugation at 13,000 g for 15 min. at 4 ºC. The supernatant was collected and the algal pellets were resuspended in the same medium. Washing was repeated five times. The first and the second
supernatants were pooled and represented the fraction of cell wall-bound acid phosphatase. The algal pellet was grown in the growing medium for 4 days at room temperature to obtain the secreted fraction of the enzyme. The secretion of the enzyme was followed according to Pfeiffer (1996).

3- Determination of growth rate of *Chlorella fusca*:

A number of criteria namely, fresh weight, content of photosynthetic pigments, cellular soluble and total phosphorus content was estimated in order to determine the specific growth of *Chlorella fusca*.

Aliquotes of 0.1 gm of *Chlorella fusca* cells were cultured for 8 days in 250 ml of a mineral medium (Grimme and Boardman 1972). Cultures were aerated and maintained continuously at light intensity of 3000 lux and room temperature (30 ± 2 °C).

The concentration of Pi in the medium was adjusted to final concentrations of 1, 15, 30 µM using NaH2PO4. Phosphorus-free medium was also used. Each treatment had three replicates. After 8 days of incubation, cells were collected by centrifugation and the fresh weight was determined as Wo – W8 where Wo is the initial weight and W8 is the weight by the end of incubation.

After fresh weight determination, cells of *Chlorella* were stored at −85 °C for subsequent determination of photosynthetic pigments, soluble proteins, intracellular phosphorus and total phosphorus as well as the activity of phosphatase enzyme.

3-1 Extraction and estimation of chlorophyll content of *Chlorella fusca*:

Previously stored algae were allowed to defrost and 0.025 gm of *Chlorella fusca* cells of each treatment were crushed thoroughly in sand and excess of acetone 80% in a mortar. The extract was centrifuged off at 13,000 rpm. The supernatant was adjusted to 10 ml by 80% acetone. The absorbance of the extract was measured at 645, 663 nm for estimation of chlorophyll a,b and total chlorophyll according to the equation reported by Arnon (1949) as follows:

- Chlorophyll a (mg/ml) = 12.7 A 663 – 2.69 A 645/ 1000. w.a x v
- Chlorophyll b (mg/ml) = 22.9 A 645 – 4.68 A663 / 1000. w.a x v
- Total chlorophyll was derived from the equation:
  - Total chlorophyll = 20.2 A 645 + 8.02 A 663 / 1000. w.a x v

Where:
A = absorbance at a given wavelength
a = length of light path in the cell (usually 1 cm)
v = volume of the extract in ml
W = fresh weight of the sample in gm

Carotenoids were estimated according to the equation reported by Metzner *et al.*, (1965) as follows:
Carotenoids = 4.2 E 452.5 – (0.0264 chl a = 0.4260 chl b) V / 1000w. Photosynthetic pigments content were also measured for non treated algae as a control.

3-2- Determination of soluble protein concentration in *Chlorella fusca*:

Determination was performed according to the method reported by Lowry *et al.*, (1951). For preparation of standard curve for protein determination, an initial concentration of Bovine serum albumin (BSA) was prepared from which a stock working standard (0.1 mg/ ml) was obtained. The working range of concentration was 1-30 mg/l. 1 ml of Lowry reagent was added to 200 µl of each concentration which was incubated at room temperature for exactly 10 min. at the end of which 100 µl of folin - cicoalteu reagent was added to each of the reaction mixtures, mixed well and incubated again for 30 min. at room temperature. Blank tubes were prepared to contain 200 µl of distilled water and the optical density was measured at 750 nm against water as a reference. The value of blank was substracted from the value of each concentration and a standard curve was constructed. Samples were treated as standards used for standard curve construction and the optical density was measured at 750 nm, which was plotted on the standard curve to obtain soluble protein concentration.

3-3- Determination of phosphorus in *Chlorella fusca* cells:

Soluble and total cellular phosphorus determinations were performed according to Moore & Chapman (1986) as follows: Stock standard solution (100 mg/L) and a working standard (8 mg/l P) were prepared. Ammonium molybdate-sulphuric acid reagent and stannous chloride reagent were prepared. The later was prepared immediately before use by dissolving 0.2 g of stannous chloride in 100 ml of 2 % (v/v) HCl. The range of concentrations of the working standard was 1.6, 3.2, 4.8, 6.4, 8.0, 9.6 and 11.2 µg / ml. To 1 ml of each standard, aliquots of 200 µl of ammonium molybdate reagent were added followed by 200 µl of stannous chloride reagent and the final volume were adjusted to 5 ml by distilled water. The reaction mixture was incubated at room temperature for 30 min. The optical density was measured at 700 nm using water as a reference. A calibration curve was constructed for the standards which were used to determine the concentration of the phosphorus in the original sample.

4- Determination of phosphatase activity in *Chlorella fusca*:

The activity of acid phosphatase (acPase) was determined according to Pfeiffer (1996) by measuring the release of P-nitrophenol (pNP) from p-nitrophenyl phosphate (pNPP). Samples of 200 µL containing enzyme were incubated with 200 µL reaction buffer containing 40 mM Mes/Tris, pH 4.5, 5 mM pNPP and 10 mM MgCl2. 6 H2O for 45 min at 30 ºC. The reaction was stopped and the color was developed by addition of 800 µl of 400 mM borate buffer, pH 9.8. The concentration of pNP was determined using a linear regression of calibration standers. All experiments were performed in ice as triplicates.
Results

As the external levels of Na2HPO4 increased, the specific growth increased (Figure 1). It is clear from this figure that all the growth criteria; the fresh weight of the alga, the total chlorophylls, the total carotenoids concentration and total protein concentrations increased by increasing the concentration of the inorganic phosphorus in the medium. Both the intracellular inorganic and the total phosphorus concentrations were also increased with increasing the external Na2HPO4 levels (Figure 2 A and B).

Figure 1. Changes in specific growth of Chlorella fusca exposed to various levels of external Na2HPO4. A: Fresh weight of the alga (mg.10^-1), B: Total protein concentration (mg / gm f.wt), C: Total chlorophyll concentration (mg / gm f.wt), D: Carotenoid concentration (mg / gm f.wt).

Figure (3) showed that changes in the specific activity of acid phosphatase were inversely related to the concentration of the external inorganic phosphorus. It is clear from this figure that addition of Na2HPO4 at
Intramolecular phosphorus concentration

Total phosphorus (μM)

Na₂HPO₄ concentration

Figure 2. Changes in inorganic and total phosphorus in *Chlorella fusca* exposed to various levels of external (Na₂HPO₄).

Figure 3. Relationship between acid phosphatase activity and concentration of inorganic phosphorus in *Chlorella fusca*.
concentrations of 1.0, 15.0 and 30.0 µM inhibited the initial activity of the enzyme by 23.0 %, 29.2 % and 38.4 %, respectively compared to control.

**Discussion**

The external level of Na2HPO4 affected the total phosphorus and the concentration of intracellular inorganic phosphorus (Pi), the lower the external Na2HPO4 the lower the intracellular Pi and total phosphorus concentration (figure 1). It has also been shown in other algae that the levels of intracellular inorganic phosphorus decreased when exposed to P-deficient conditions (Barret-Lennard et al., 1982; Dracup et al., 1984; Lundberg et al., 1989; Tillberg and Rowley, 1989; Duff et al., 1994; Lee, 2000).

The present results demonstrated a linear correlation between the activity of acid phosphatase, intracellular inorganic phosphorus and total phosphorus concentrations referring to that in *Chlorella fusca* the acid phosphatase can be used as an indicator of P-deficiency. The increment in acid phosphatase activity in Pi–limited conditions suggests that the acid phosphatase could be involved in polyphosphate degradation in the tested alga (*Chlorella fusca*) as has been suggested for some higher plants such as soybean vegetative storage proteins (Dewald et al., 1992) and some ornamental plants (Duff et al., 1994). Acid phosphatase activity also has been used in higher plants as a biochemical marker of P-limitation (Barret-Lennard et al., 1982) who worked on wheat leaves and Ibrahim (2005) who worked on pollen grains of *Zea mays*. Increased the activity of acid phosphatase in response to P-deficient conditions and the secretion of the enzyme have been reported in some crop plants which were studied by Tadano & Sakai (1991). Similarly, in accordance with data of the present investigation, Gilbert et al. (1999) indicated that P-deficiency in white lupin roots (*Lupinus albus*) induced a number of coordinated metabolic changes in roots, and possibly in the whole plant.

Similarly to the results obtained by El-Shahed et al. (2006), the in-vitro experiments in this study showed that phosphate ions are potent inhibitors of the activities of the acid phosphatase enzyme in *Chlorella fusca* (figure 3). A high correlation with intracellular Pi concentration was found for acid phosphatase activity. The induction of acid phosphatase activity is therefore possibly due to a lower intracellular Pi concentration. This study also showed that growth of *Chlorella fusca* depends on the external supply of Na2HPO4 and that there is a linear relationship between external Na2HPO4 level and the specific growth rates and hence there is a reverse relationship between growth rates and the secretion or even the activity of the enzyme (data not shown) and this observation is in contrast with the conclusion obtained by Ibrahim et al. (2002) who worked on acid phosphatase produced by germinating lily pollen grains.
References


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