

## PLANKTON COMMUNITY, WATER QUALITY AND GROWTH PERFORMANCE OF NILE TILAPIA AS AFFECTED BY HUMIC ACID IN AQUACULTURE PONDS

Mohamed G. Battah<sup>1</sup>, Aida M. Dawah<sup>2</sup>, Ayman A. Ammar<sup>2</sup> and Mohamed M. Salama<sup>2\*</sup>

<sup>1</sup>. Botany department, Faculty of Science, Banha University, Egypt

<sup>2</sup> Central Laboratory for Aquaculture Research (CLAR), Abbassa, Abou-Hammad, Sharkia, Egypt.

### Abstract

This study was carried out at Wady El-Ryan, Fayoum Governorate, Egypt to evaluate the influence of humic acid on enrichment of plankton community, water quality, growth performance and pond productivity of Nile tilapia. Eight earthen ponds of a total area of two feddan were used. These fish ponds were divided into four treatments; two replicates per each as the first treatment ponds (T1 and T2) were stocked with Nile tilapia fry 14000 fish/Fadden. The second ponds (T3 and T4) were stocked with 28000 fish/Fadden. T2 and T4 treatment ponds received only supplemental feed at a rate of 3% of body weight (bw), while T1 and T3 were treated with humic acid at a rate of 3 Kg/Feddan during the preparation of the ponds, then they received a supplemental feed adding on it 1.5 Kg of humic acid/ ton feed. Comparison between treatments showed that there were no significant differences ( $p > 0.05$ ) in water temperature, salinity, total hardness and total alkalinity. Dissolved oxygen, orthophosphate, Chlorophyll "a", concentrations in T1 and T3 ponds were significantly higher ( $p < 0.05$ ) than those of T2 and T4 ponds. Free ammonia, nitrate and nitrite concentrations in T2 and T4 ponds were significantly higher ( $p < 0.05$ ) than those of T1 and T3 ponds. Secchi disk reading was significantly higher in T4 ponds than that of the other treatment ponds. Phytoplankton and zooplankton abundances were significantly higher in T1 and T3 ponds than those of T2 and T4. The final weight of Nile tilapia was maximized at T1 ponds, while the lowest one was in T4 ponds.

**Keywords:** Humic acid; Nile tilapia; Water Quality; Phytoplankton; Zooplankton.

### Introduction

Humic acid is one of the major components of humic substances, which are dark brown and major constituents of soil organic matter. Humic substances are excellent natural and organic way to provide a concentrated dose of essential critical nutrients, vitamins and trace elements not only to stabilize the plankton blooms in aquaculture ponds but also to improve water quality. The humic acid

(ISSN: 1110-8649)

alone is suitable for freshwater algal production and with better water quality variables (**Bakhsh, 2001**). Besides, humic acid has the capability of reducing the negative effect of high dosage of nitrogen (N) and phosphorus (P) in treatments. Humic acid acts as supplier and storehouse for N and P for algae and phytoplankton in aquatic ecosystem. Some studies have also shown that humic acid can increase the productivity of phytoplankton (**Vrana and Votruba, 1995**) in environment and laboratory experiments. Humic substances are may eventually prove useful for controlling the growth of noxious cyanobacteria in water environments (**Sun *et al.*, 2006**). Using of fulvic acid with certain dose improves production performance of tilapia and it has significant differences ( $p < 0.05$ ) for both final body length and weight (**Wet and Visagie, 2010**). The fish exposed to humic acids were significantly longer and heavier than the control group (**Meinelt *et. al*, 2003**) and they added that all the treatment groups exposed to humic acids continued steady growth after stress (2 weeks of daily nettings) . Dietary humic acid supplementation during growing period at certain dose rate can be used to improve body weight gain and feed efficiency (**Avic *et al.*, 2007**). The aim of this study was to investigate the potential use of humic acid in improving productivity (plankton) and provide more insight on water quality variables in a freshwater aquaculture, Suitability of pond water for fish production.

### ***Materials and Methods***

The present study was conducted in eight earthen ponds 8400m<sup>2</sup> surface areas each, 150 cm water depth. The ponds were prepared by drying for about two weeks before the beginning of the experiment. Ponds were located at Wadi EL-Ryan, Fayoum Governorate, Egypt. Ponds were filled with water from drainage canal. The ponds were divided into two groups; first group with humic acid as 3 Kg/ Faddan before stocking with fish and second group without humic acid

Humic acid in the experiment was procured from a Turkish company by an Egyptian sole agent (IBN ELWALED CO.) as commercial product *HUMAPOL* fertilizer which consists of Humic and Fulvic acid, Manganese, Zinc, Iron, Copper, Cobalt, Iodine Selenium, and Koalin, 850, 5.00, 6.00, 60, 5.00, 0.20, 1.00, 0.17, and 2.00 gm/kg respectively as the company report.

The groups of the experiment divided as:

Tretments	Group one		Group two	
	(T1)	(T2)	(T3)	(T4)
Number of Fish/faddan	14000	14000	28000	28000
<i>Humapol</i>	present	without	present	without

These treatments were assigned to ponds at random in a completely randomized block design and each was replicates two times. Ponds were stocked with Nile tilapia (*Oreochromis niloticus*) with initial weight was 10 g/ fish. The experimental period was 24 weeks.

Fish samples were taken monthly about 200 fish/ pond for calculating the growth performance and the feed quantity per pond, the pellets of feed 27 % protein was used throughout the experiment and was provided twice daily five days/week at a rate of 3 % of body weight for fish in each pond.

#### **Water quality**

Temperature (°C) and dissolved oxygen (DO mg/L) were measured by oxygen meter (Aqualytic OX24). Salinity (g/L) and pH were measured bi-weekly using pH/conductivity meter (Orion 543). Secchi disk (SD cm) visibilities, ammonia (mg/L), nitrite (mg/ L), nitrate (mg/ L), total alkalinity as CaCO<sub>3</sub> (mg/L), total hardness (mg/L), orthophosphate (mg/L) and Chlorophyll "a" (µg/L) were also measured once a two week using standard methods (APHA, 2000).

#### **Phytoplankton**

Phytoplankton in the sample was concentrated by settling 500 ml sample in a volumetric cylinder for about 24 hours after being preserved in lugol's solution (prepared by dissolving 20gm of potassium iodine (KI) and 10gm of iodine crystal in 200 ml distilled water solution containing 20 ml glacial acetic acid), at a ratio 0.3 ml lugol's solution to 50 ml sediment (APHA, 1985). The surface water was siphoned and the sediment was examined. One ml of sample was transferred into Sedgwick-Rafter cell and counted microscopically. Three replicates of each sample were investigated. Different algal species were identified according to (Prescott, 1962 and 1978). Phytoplankton cells were identified to four divisions, which are: green algae (Chlorophyceae), blue-green algae (Cyanobacteria), diatoms (Bacillariophyceae), and euglena (Euglenophyceae) and different species were identified and counted. All colonial, filamentous and unicellular organisms were counted as one unit (cell). The equation was used to determine phytoplankton diversity according to (Shannon and Weaver, 1949).

**Zooplankton:** Water sample of 10 L was collected from each pond by 20µm pore size plankton net. The samples were measured biweekly during the experiment.

**Growth performance:** Growth performance was determined and feed utilization was calculated as follows:

**Weight gain (g) =  $Wt_2 - Wt_1$** ; where  $Wt_1$  and  $Wt_2$  are the initial and final fish weight, respectively.

**Daily Weight Gain (DWG)** was calculated as:  $(Wt_2 - Wt_1) / t$

Where:

$Wt_1$  = initial fish weight in grams.

$Wt_2$  = final fish weight in grams.

$t$  = period in day.

**Specific growth rate (SGR)** =  $(\ln wt_2 - \ln Wt_1) \times 100 / t$ .

Where:

$\ln = (\log 10x)^{3.303}$

$t$  = is the number of days in the feeding period.

**Food conversion ratio (FCR):** The food conversion ratio was calculated according to following equation:

$$FCR = \frac{\text{Total feed consumption (g)}}{\text{Final body weight (g)} - \text{initial body weight (g)}}$$

### Statistical analysis

Statistical analysis was performed using the analysis of variance (ANOVA) and Duncan's multiple range tests to determine differences among treatments. All statistics were carried out by using Statistical Analysis Systems (SAS) program (SAS, 2000).

### Results and Discussion

Bi-weekly monitored water quality parameters are summarized in Table (1). All parameters were within an acceptable range for tilapia culture under Egyptian conditions (Shaker, 2008). There were significant differences in water quality parameter between treatments with and without humic acid fertilizer treatment ponds; and different stocking density. The pH values were 8.5; 9.4; 8.6 and 9.2 for T1; T2; T3 and T4 respectively. This variation could be explained by the photosynthetic uptake of  $CO_2$  and bicarbonate that substituted hydroxyl ions. These results indicated that the using of humic acid fertilizer and increased fish density were effected significantly on pH values (Shaker and Mahmoud, 2007). Temperature ranged from 28.4 to 28.8°C in the treatments during the whole period. Secchi disc reading was significantly higher ( $P < 0.05$ ) in T2 and T4 than in T1 and T3 treatments. These results showed low productivity of water in the T2 and T4 treatment than other treatments. Also, the SD was significantly decreased ( $P < 0.05$ ) in humic acid fertilizer treatments than without humic acid fertilizer treatments. These results are agreed with (Sa'nchez-marin and Beiras, 2011)

who found that addition of humic acid fertilizer stimulates the growth of phytoplankton and zooplankton causing low water transparency in reading. Dissolved oxygen concentration was significantly ( $P < 0.05$ ) higher in T1 and T3 treatments than in the other treatments Table (1). These results may be due to the increase of photosynthetic led to increasing of DO production by phytoplankton (**Shaker, 2008**). The concentration of nitrate and phosphate showed significance higher ( $P < 0.05$ ) values in humic acid treatments through the experimental period.

The pH, temperature and dissolved oxygen were the most influencing parameters in fish ponds, where their values in all ponds, although fluctuated from time to time, they stayed within the acceptable and favorable levels required for growth, survival and well being of the tested fish species (**Shaker et al., 2013**). The decrease of DO concentrations in treatment without humic acid fertilizer may have resulted from the lower grazing pressure by fish, which resulted in larger planktonic communities, subsequently contributing to higher accumulation rates of organic material on the bottom, leading to bacterial decomposition and increased sediment oxygen consumption ( **Smith and Piedrahita, 1988**). Elevated total ammonia concentrations may indicate to higher sediment organic matter accumulation and decomposition rates in these treatments. However, DO levels were always within the optimum range for different fish species in polyculture production in the earthen ponds (**Shaker et al., 2009**). DO increases feed consumption, feed efficiency, metabolism, and growth of fish (**Azim et al., 2005 and Shaker, 2008**). Numerous studies have shown increases in oxygen consumption after feeding due to the metabolic cost of digesting and assimilating the nutrients from the feed. The concentration of DO in water is dramatically affected by stocking density, water temperature, water flow and pond management (**Shaker and Abdel Aal, 2006; Shaker and Mahmoud, 2007 and Shaker et al., 2009**).

Water quality measurements made during the trial indicated that there were statistically significant differences among treatments as a result of the presence of the humic acid fertilizer. There was a trend of higher Secchi disc values for of the presence of humic acid, including phytoplankton, in the water column and lower chlorophyll-a concentrations in water samples from ponds with humic acid, indications that there may be a trade-off between phytoplankton and humic acid activity and phytoplankton production.

The data in Table (1) show the average values of nitrogen compounds in water under different treatments. The average values of  $\text{NH}_3$  were 0.22; 0.59; 0.16; and 0.44mg/l for T1; T2; T3 and T4 respectively. The average values of  $\text{NO}_2$  were 0.078; 0.148; 0.06and 0.1mg/l for the same treatments respectively.

**Table (1): Means ( $\pm$ ) standard errors of some physico-chemical parameters in earthen ponds under different treatments during the experimental period**

Item Treat.	Temp °C	pH	SD cm	DO mg/l	NH <sub>4</sub> -N mg/l	NH <sub>3</sub> -N mg/l	NO <sub>2</sub> -N mg/l	NO <sub>3</sub> -N mg/l	Total alkalini ty mg/l	Total hardnes s mg/l	Salinit y g/l	Ortho phosph h. mg/l	Chlor ophyll -a µg/l
T1	28.4 <sup>a</sup> $\pm$ 2.5	8.5 <sup>b</sup> $\pm$ 0.5	10 <sup>b</sup> $\pm$ 1.2	7.6 <sup>a</sup> $\pm$ 0.3	1.08 <sup>b</sup> $\pm$ 0.1	0.22 <sup>b</sup> $\pm$ 0.02	0.078 <sup>b</sup> $\pm$ 0.002	0.164 <sup>b</sup> $\pm$ 0.01	294 <sup>a</sup> $\pm$ 25.00	556 <sup>a</sup> $\pm$ 20.00	2.2 <sup>a</sup> $\pm$ 0.10	0.31 <sup>a</sup> $\pm$ 0.02	88.6 <sup>b</sup> $\pm$ 6
T2	28.8 <sup>a</sup> $\pm$ 2.0	9.4 <sup>a</sup> $\pm$ 0.5	14.0 <sup>a</sup> $\pm$ 0.5	4.2 <sup>c</sup> $\pm$ 0.4	1.84 <sup>a</sup> $\pm$ 0.2	0.59 <sup>a</sup> $\pm$ 0.04	0.148 <sup>a</sup> $\pm$ 0.01 <sup>a</sup>	0.296 <sup>a</sup> $\pm$ 0.02	278 <sup>a</sup> $\pm$ 25.00	468 <sup>a</sup> $\pm$ 25.00	2.3 <sup>a</sup> $\pm$ 0.10	0.21 <sup>b</sup> $\pm$ 0.04	54.6 <sup>a</sup> $\pm$ 4
T3	28.4 <sup>a</sup> $\pm$ 2.5	8.6 <sup>b</sup> $\pm$ 0.5	11.0 <sup>b</sup> $\pm$ 0.5	6.2 <sup>b</sup> $\pm$ 0.4	0.92 <sup>b</sup> $\pm$ 0.1	0.16 <sup>b</sup> $\pm$ 0.01	0.06 <sup>b</sup> $\pm$ 0.001	0.120 <sup>b</sup> $\pm$ 0.02	288 <sup>a</sup> $\pm$ 24.00	548 <sup>a</sup> $\pm$ 22.00	2.3 <sup>a</sup> $\pm$ 0.10	0.32 <sup>a</sup> $\pm$ 0.02	94.3 <sup>b</sup> $\pm$ 8.5
T4	28.8 <sup>a</sup> $\pm$ 2.0	9.2 <sup>a</sup> $\pm$ 0.4	16 <sup>a</sup> $\pm$ 0.5	3.4 <sup>c</sup> $\pm$ 0.5	1.44 <sup>a</sup> $\pm$ 0.12	0.44 <sup>a</sup> $\pm$ 0.03	0.10 <sup>ab</sup> $\pm$ 0.01	0.230 <sup>ab</sup> $\pm$ 0.02	286 <sup>a</sup> $\pm$ 25.00	556 <sup>a</sup> $\pm$ 22.00	2.2 <sup>a</sup> $\pm$ 0.10	0.15 <sup>b</sup> $\pm$ 0.02	44.7 <sup>a</sup> $\pm$ 4.2

The increase of  $\text{NH}_3$  in the treatments without humic acid could be explained by the decomposition of organic matter and via the direct excretion of ammonia by the large biomass of fish. These finding indicated that the humic acid improved water quality. The  $\text{NO}_2$  and  $\text{NO}_3$  concentrations in water followed the same trend of ammonia-nitrogen. These results may be due to the consumption of nitrate (which is an essential nutrient) by phytoplankton communities. Also, the increase of nitrate in T2 and T4 may be related to the increase of phytoplankton standing corps. There a positive correlation between nitrate content and total phytoplankton which may be attributed to high consumption rate of  $\text{NO}_3$  by the algae. These results are in harmony with those obtained by (Shaker *et al.*, 2009). Water quality is improved through the higher rate of nitrification and ammonia uptake by phytoplankton algae (Thompson *et al.*, 2002 and Shaker *et al.*, 2013). The uneaten feed and dead periphyton cells remain attached with the surface sediment, providing a rich source of organic nutrients for heterotrophs associated with periphyton layer. Processing of this organic matter yields inorganic nutrients that can be utilized by living algae again. However, there is very little opportunity for humic acid treatments to grow in well managed fish culture.

Phosphorus is an element that, in its different forms, stimulates the growth of aquatic macrophytes and algae in water bodies. The average values of dissolved phosphorus in the same treatments were 0.31; 0.21; 0.32 and 0.15mg/l, respectively Table (1). These results clear that the concentration of available phosphorus were significantly decrease ( $P < 0.05$ ) in T2 and T4 treatments. These results may be due to the added of humic acid led to release of phosphorus from the surface layer of soil to water (Singh and Amberger, 1991). The chlorophyll a concentrations in water followed the same trend of phosphorus.

Humic acid assemblages lead to increased retention of nutrients through firstly; it can remove nutrients from the water column and cause a net flux of nutrients toward the sediments (Singh and Amberger, 1991). Second, it can slow water exchange across the sediment/water column boundary thus decreasing advective transport of P away from sediments (Sarir *et al.*, 2006). Third, they can intercept nutrients diffusing from the benthic sediments or senescent macrophytes. It can cause biochemical conditions that favor P deposition.

The presented data in Table (1) show the average values of salinity, total alkalinity and total hardness were not significance ( $P < 0.05$ ) changed in all treatments during the experimental period. These results indicated that the added of humic acid did not affect on salinity, total hardness and total alkalinity.

#### **Plankton algae development**

It has been found that humic acids in the water can affect the development of plankton algae. By the addition of humic acids fertilizer in the water, the development of Chlorophyta and Bacillariophyta was stimulated, while

the numbers of Cyanophyta in T1 and T3 were lower mainly the development was inhibited than those registered in the other variation T2 and T4. Abundance of phytoplankton was positively affected by HA treatments ( $r = 0.578$ ;  $P < 0.01$ ), and it was significantly higher at T1 and T3 than that of T2 and T4 **Table (3)**.

The maximum number of phytoplanktons was recorded in October T1 and at T3 (181 and 129 org. /ml), respectively while the minimum value was recorded in May and June at T4 (38 and 45 org. /ml respectively). **Table (2)** shows the different species of each phytoplankton division. The recorded species for each division were 21, 20, 17 and 4 for Bacillariophyceae, Chlorophyceae, Cyanobacteria and Euglenophyceae respectively. Phytoplankton of T2 and T4 was characterized by a predominance of blue-green algae in terms of their numbers. *Merismopedia elegans* and *Phormidium* sp dominated. *Microcystis aeruginosa* and *Chroococcus minutus* were subdominant.

Other species such as *Lyngbya birgei*, *Gleocapsa* sp., and *Oscillatoria hamelii* were also present in the ponds. Chlorophyceae such as *Oocystis* sp., *Scenedesmus* sp and *Tetraedron* sp were found to be the most abundant species. In addition, *Ankistrodesmus*, *Chlorella*, *Closterium* sp. and *Cosmorim* sp. were recorded. The maximum *Scenedesmus* number was recorded in August in T1 (19 org. /ml), while the minimum number was obtained in May in T4 (3 org. /ml). The highest number of *Dictyosphaerium* sp. was recorded in August in T3 (8 org. /ml).

The most abundant species of Bacillariophyceae were *Navicula* sp., *Nitzschia* sp., and *Synedra* sp. Other species of Bacillariophyceae were *Cyclotella* sp., *Cymbella* sp., *Diatoma* sp., *Gemphonema* sp., and *Pinnularia* sp., *Navicula* sp. and *Nitzschia* sp. were the most abundant genera at all treatments during the study period. *Navicula* sp. exhibited the highest number in September and October at T1, (10 and 12 org. /ml, respectively). The highest number of *Nitzschia* sp. was recorded in October at T1 (10 org. /ml) and at T3 (9 org. /ml). Moreover, *Synedra* sp. was abundant at T1 and T2, and the highest number was obtained in October in T1 (16 org. /ml). Euglenophyceae was represented only by the genera *Euglena* and *Phacus*. They were not recorded in all treatments during the experimental period. Generally, *Phacus* was less abundant than *Euglena*. The high occurrence of *Euglena* sp. was recorded during May in T4 and August in T1 (4 and 4 org. /ml, respectively). In case of *Phacus*, the highest number was obtained in August in T4 and T1 (2 and 3 org. /ml, respectively). During May, diatoms appeared as the most abundant group in T1 with variable contributions at each treatment. On the other hand, the green alga species followed the dominant diatoms in abundance during May. In June, the contributions of the algal groups were approximately similar to that found in May. During the period of June Cyanophyceae was constituted a large part of phytoplankton and Euglenophyceae was the least, Fig. (1).

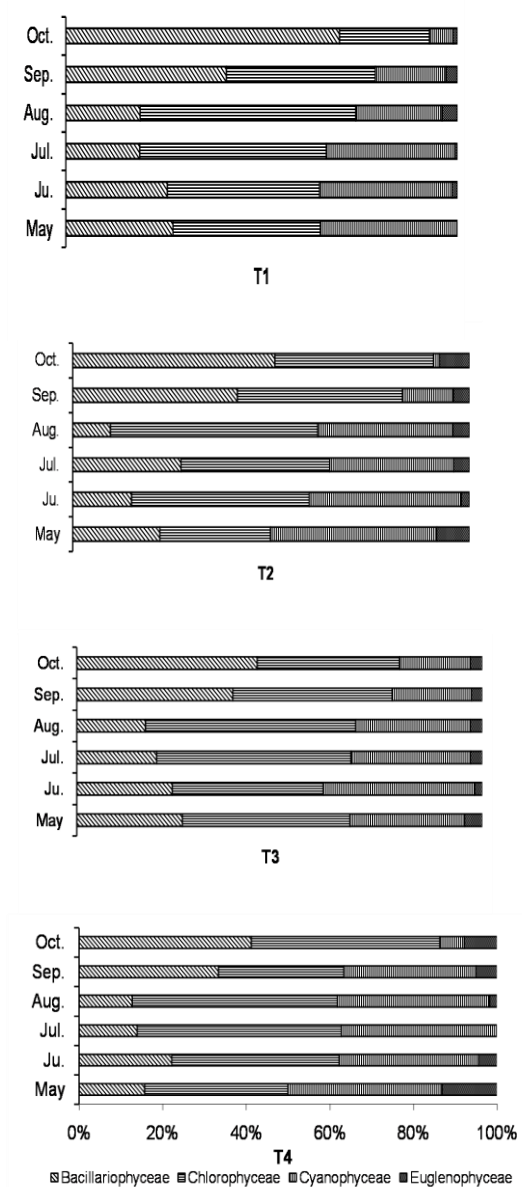


Table 2. Monthly variation in phytoplankton species in earthen ponds stocked with Nile tilapia and subjected to humic acid fertilizer

Phytoplankton species	T1						T2						T3						T4					
	May	June	July	Aug.	Sept	Oct.	May	June	July	Aug.	Sept	Oct.	May	June	July	Aug.	Sept	Oct.	May	June	July	Aug.	Sept	Oct.
Bacillariophyceae																								
<i>Amphora ovalis</i>	—	—	1	1	—	—	—	—	—	1	—	—	1	—	—	1	—	—	—	—	—	—	2	1
<i>Cyclotella meneghiniana</i>	1	1	4	2	3	4	1	—	2	—	2	3	—	—	—	—	3	3	—	—	—	—	2	1
<i>Cymbella affinis</i>	2	1	1	2	4	5	—	—	—	1	1	—	—	—	1	—	2	3	—	—	—	—	1	2
<i>C. silesiaca</i>	—	1	—	1	4	5	2	—	2	2	2	1	—	—	1	—	4	—	—	—	—	1	2	—
<i>Cymbella sp.</i>	1	—	—	—	4	4	—	—	—	1	1	—	—	—	2	1	4	1	2	—	—	1	—	2
<i>Diatoma mesodon</i>	—	1	1	2	4	6	—	—	2	—	3	2	—	—	1	—	4	3	—	1	1	1	1	2
<i>D. vulgaris</i>	—	1	3	3	4	6	—	2	—	—	2	1	1	—	—	—	4	6	1	—	—	—	2	—
<i>Fragilaria capucina</i>	1	2	—	2	2	2	1	—	—	—	—	2	—	—	—	2	2	2	—	—	—	—	2	—
<i>Gomphonema gracile</i>	1	1	2	3	3	5	—	—	2	—	2	2	—	3	—	1	3	5	1	—	—	1	1	—
<i>G. olivaceum</i>	—	—	—	2	2	—	—	—	—	2	3	3	1	1	2	1	2	—	1	2	—	—	2	1
<i>Gomphonema sp.</i>	1	—	4	3	7	8	1	—	—	—	4	3	—	—	2	2	4	5	1	1	—	—	—	—
<i>Navicula radiosa</i>	—	2	3	2	6	7	—	—	2	—	1	2	1	2	1	2	5	7	—	—	2	1	—	—
<i>Navicula sp.</i>	2	1	1	2	4	5	—	—	—	—	2	1	2	1	1	3	4	5	—	3	2	—	1	2
<i>Nitzschia alpina</i>	—	—	—	—	3	3	—	2	—	—	1	1	—	2	2	1	2	3	1	1	—	—	1	1
<i>N. closterium</i>	1	—	1	1	3	4	—	—	—	2	—	2	2	2	1	2	2	3	—	—	—	—	—	1
<i>N. palea</i>	—	—	—	—	2	1	1	2	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—
<i>N. radicola</i>	—	2	—	—	—	2	—	—	—	—	—	—	—	—	—	—	—	2	—	—	—	—	—	1
<i>Pinnularia sp.</i>	1	2	—	—	1	1	—	—	2	—	—	1	—	—	—	—	—	1	—	—	—	—	—	—
<i>Synedra acus</i>	2	2	3	2	4	5	1	—	—	—	2	3	—	—	—	—	2	5	—	—	—	—	1	2
<i>Synedra ulna</i>	—	—	—	—	2	3	—	—	2	—	1	3	—	—	—	—	1	3	—	—	—	—	1	1
<i>Synedra sp.</i>	—	1	—	—	6	8	1	1	—	—	3	2	1	1	3	—	4	4	—	—	—	2	2	2
Chlorophyceae																								
<i>Ankistrodesmus braunii</i>	1	2	3	5	—	—	—	2	1	1	—	—	—	—	2	—	3	—	—	—	—	2	—	—
<i>A. convolutus</i>	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	—	—
<i>A. falcatus</i>	—	—	1	1	—	—	—	—	—	—	—	—	1	2	—	—	—	2	—	—	—	—	—	—
<i>A. setigera</i>	—	—	2	—	—	2	—	—	—	—	—	—	—	—	—	—	3	—	—	—	—	—	—	—
<i>Ankistrodesmus sp.</i>	—	2	5	8	1	—	—	2	1	2	—	1	2	3	3	5	7	9	1	1	3	—	6	—
<i>Chlorella vulgaris</i>	3	4	11	11	13	17	5	2	2	5	8	13	3	1	8	11	14	9	3	8	6	4	6	8
<i>Closterium sp.</i>	2	—	2	4	2	—	—	2	—	2	—	—	3	3	—	3	—	5	—	—	2	2	2	3
<i>Cosmorium sp.</i>	—	—	—	2	—	—	—	1	—	—	—	—	1	—	—	—	—	—	—	—	—	—	2	—
<i>Dictyosphaerium sp</i>	—	—	—	3	—	—	—	—	—	—	3	—	—	2	5	8	5	—	—	—	—	2	—	—
<i>Gomphoshaeria sp</i>	1	—	—	4	—	—	—	—	1	—	—	—	1	—	3	5	3	3	—	—	—	1	—	1
<i>Oocystis limneticus</i>	—	1	5	7	10	8	—	2	4	3	—	1	1	2	—	2	3	2	2	2	—	2	—	—
<i>Oocystis solitarius</i>	1	1	6	9	6	4	—	—	—	—	—	—	—	—	—	—	2	—	—	—	—	—	—	—
<i>Oocystis sp.</i>	2	3	11	8	6	6	2	—	1	5	5	—	1	1	3	2	1	3	—	—	5	5	—	4
<i>Scenedesmus acuminatus</i>	3	5	9	17	6	3	3	3	3	8	11	9	2	2	6	5	7	—	3	5	4	4	—	—
<i>S. quadricauda</i>	—	—	1	—	1	—	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	—	—
<i>Scenedesmus sp.</i>	—	2	—	2	—	—	—	—	—	2	—	—	—	—	—	3	—	2	—	—	—	—	—	—
<i>Selenastrum sp.</i>	1	2	—	3	3	—	—	—	1	3	—	—	1	—	—	2	—	2	2	—	—	1	—	2
<i>Tetradron caudatum</i>	—	—	—	1	1	—	—	—	—	1	—	—	—	—	—	1	—	—	—	—	—	—	—	—
<i>Tetradron minimum</i>	—	—	2	2	3	—	—	2	2	1	—	1	2	2	2	1	—	1	—	—	—	2	1	4
<i>Tetradron sp.</i>	4	5	4	8	11	2	—	5	3	5	3	1	1	1	3	2	1	1	2	2	1	2	1	1
Cyanobacteria																								
<i>Anacystis sp.</i>	—	4	—	2	—	—	—	—	3	—	1	—	2	—	—	2	—	3	—	—	1	3	2	—
<i>Chroococcus minutus</i>	2	6	13	7	9	7	2	1	—	2	—	1	3	—	1	4	5	2	2	8	6	8	7	3
<i>Chroococcus sp.</i>	1	2	3	—	—	—	2	—	—	2	—	—	1	4	3	—	3	1	3	2	—	—	—	—
<i>Gleocapsa sp.</i>	—	—	—	4	—	—	1	—	—	—	—	—	1	—	2	—	—	2	2	—	—	—	2	1
<i>Lyngbya birgei</i>	1	1	—	1	—	—	—	2	—	—	—	—	—	—	—	5	—	—	—	1	—	—	1	1
<i>Merismopedia glauca</i>	—	—	2	—	4	—	2	7	3	—	—	—	—	5	—	2	—	—	—	—	3	2	—	—
<i>M. punctata</i>	3	—	—	—	2	—	—	—	—	2	—	—	—	2	—	2	—	—	—	—	—	—	—	—
<i>M. tenuissima</i>	—	—	2	—	—	—	—	—	—	3	—	—	—	1	—	3	—	1	—	2	—	—	1	2
<i>Merismopedia sp.</i>	2	—	6	8	—	—	2	—	2	—	—	—	2	2	—	—	3	—	—	—	3	—	—	—
<i>Microcystis aeruginosa</i>	—	—	2	—	3	—	—	3	3	3	2	—	1	1	2	1	1	1	2	2	2	1	2	1
<i>Microcystis flos-aquae</i>	—	2	—	—	3	—	—	—	—	2	—	—	—	—	—	1	—	—	—	—	—	1	—	—
<i>Microcystis sp.</i>	2	—	—	—	—	1	1	1	1	3	—	—	—	—	2	—	1	1	2	—	—	1	—	—
<i>Oscillatoria hamelii</i>	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	—	—
<i>O. rubescens</i>	—	—	—	—	—	—	1	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	—	—
<i>O. tenuis</i>	2	2	—	3	—	—	2	1	—	2	—	—	1	—	2	—	3	2	—	—	—	1	—	—
<i>phormidium sp.</i>	4	7	11	14	8	3	3	—	4	6	3	—	2	5	9	7	5	5	2	—	3	—	2	1
Euglenophyceae																								
<i>Euglena acus</i>	—	—	1	2	1	—	1	—	—	1	2	3	—	1	2	1	2	—	1	—	—	1	2	—
<i>Euglena spp.</i>	—	—	—	2	2	2	1	1	1	—	—	—	1	—	—	2	—	1	3	2	—	—	1	3
<i>Phacus longicauda</i>	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	—	—	—	—	—	—	—	—
<i>Phacus sp.</i>	—	1	—	3	2	—	1	—	1	1	1	2	1	—	—	—	1	2	1	—	—	—	—	—

**Table 3** Variance and distribution of major phytoplankton groups (org/ml) in earthen ponds stocked with Nile tilapia and Subjected to humic acid fertilizer

phytoplankton Groups	T1					T2					T3					T4								
	May	June	July	Aug	Sept	Oct.	May	June	July	Aug	Sept	Oct.	May	June	July	Aug	Sept	Oct.	May	June	July	Aug	Sept	Oct.
Bacillariophyceae	13	18	24	33	68	126	8	7	14	7	30	33	12	12	14	18	45	69	6	10	6	7	20	21
%	31.7	26	19	19	41.2	70	22.2	15	27	9.5	41.6	51	26	23.5	20	17	38.4	35	16	22.2	14	13	33.3	41.1
Chlorophyceae	18	27	60	95	63	42	10	21	19	38	30	26	19	19	34	55	46	39	13	18	21	27	18	23
%	43.9	39	48	55	38.1	23	27.7	45	37	52	41.6	40	41.3	37.2	48	52	39.3	28	34	40	49	49	30	45
Cyanophyceae	17	24	41	39	29	11	15	18	16	25	9	1	13	19	21	30	23	18	14	15	16	20	19	3
%	41.4	34	33	22	17.5	6.1	41.6	38	31	34	12.5	1.5	28.2	37.2	30	28	19.6	14	37	33.3	37	36	31.6	5.8
Euglenophyceae	—	1	1	7	5	2	3	1	2	3	3	5	2	1	2	3	3	3	5	2	—	1	3	4
%	0	1.4	0.8	4	3.03	1.1	8.3	2.1	3.9	4.1	4.1	7.6	4.3	1.9	2.8	2.8	2.5	2.3	13	4.4	0	1.8	5	7.8
Total number	41	70	126	174	165	181	36	47	51	73	72	65	46	51	71	106	117	129	38	45	43	55	60	51
%	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100



**Fig. 1. Monthly variations in percentage composition of different phytoplankton groups in the different treatments (T1, T2, T3 and T4) during the experimental period.**

In August, green algae became again the predominant (52-57%), while diatoms mostly ranked the second abundant group with two exceptions at T1 and T3. During the first half of September, green algae were still dominant in the whole area, replaced alternatively in late September and October by diatoms and blue-green algae, Fig. (1).

The T1 and T3 ponds exhibited high number of phytoplankton organisms in comparison to that of T2 and T4. This result might be was proportional to the daily rate of humic acid addition and that was similar to (Carlsson *et al*, 1995) who found that addition of humic acid stimulates bacterial, phytoplankton and ciliated growth in coastal waters and that humic bound nitrogen is to some extent available for phytoplankton. Humic substances may have something to do in the improvement of the photosynthetic activity and efficiency observed, through a direct and/ or indirect stimulatory effect on the algal community, (Conzonno and Cirelli, 1996). The number of phytoplankton depended mainly on the number of Cyanophytceae and Chlorophyceae, which were the most abundant groups ( $r = 0.956$  and  $0.931$ , respectively;  $P < 0.05$ ), while Bacillariophyceae and Euglenophyceae were the less abundant ones in all treatments ( $r = 0.605$  and  $0.189$ , respectively;  $P < 0.05$ ). The sensitivity of Cyanobacteria to humic acid may also explain patterns in the field. Thus, ponds which were treated with humic acid do not support the Cyanobacterial blooms; the opposite of poor humic ponds (T2 and T4). Thus, Cyanobacteria appear to be unable to use their accessory pigments (particularly phycocyanin) to exploit the reddish light prevailing in humic-rich lakes (Keskitalo and Eloranta, 1998). Addition of humic acids has a large effect on phytoplankton composition, since it stimulates the growth of chlorophyceae and Bacillariophyceae while it has negative effect on Cyanophyta, (Klochenko *et al*, 2011). High numbers of Bacillariophyceae (diatoms) were presented in treated ponds which were characterized by daily addition of HA have than T2 & T4 since humic acid has a stimulatory action on growth of diatoms (Prakash *et.al*, 1973). Spore formation of Bacillariophyceae can be induced by high temperature ( $24^{\circ}\text{C}$ ) whereas low temperatures defer it (Adolf *et al.*, 2009); this explains the lowest number of Bacillariophyceae during high temperature period. Euglenophyceae increase coincides with high eutrophication levels, (Taniguchi *et al.*, 2005); while humic substances prevent and make remediation of eutrophication (Steinberg *et al*, 2008).

#### Zooplankton Abundance

The common zooplankton groups were Cladocera, Copepods and Rotifers in all treatments. Rotifer was dominant zooplankton group, while Copepods, Cladocera and Ostracoda exhibited sub-abundance in all treatments Table (4). Zooplankton abundance was positively affected by HA treatments ( $r = 0.420$ ;  $P < 0.01$ ) and it was greater in T1 and T3 than that of T2 and T4 ( $P < 0.05$ ). The maximum number of zooplankton was obtained in October at T1 (87 org. /L)

followed that in T3 (86 org/L). The minimum number of zooplankton of 23 org./L was recorded in T2 and 19 org./L in T4 ponds in May Table (4). Dominant Cladocera included *Moina*, *Bosmania*, *Diaphanosoma* and *Daphnia*. Cyclops was the most common copepod and common Rotifers included *Brachionus* and *Keratella*. *Cypris* was the most common of Ostracoda.

A positive correlation between the content of the humic substances and the total number of zooplankton was found in T1 and T3. (Gutseit *et al.*, 2007) showed that there are differences in food quality between humic and clear waters that affect zooplankton growth and reproduction. The large influence of humic substances can have bearings on food quality for zooplankton, since it can affect, for instance, the relative contribution of algal (high food quality), non-algal and detrital (low food quality) carbon to the total organic carbon pool, as well as microplankton community structure and composition (Hessen *et al.*, 1989). In treated humic water, rotifers appear to predominate in the zooplankton (Kankaala, 1988), although in some cases cladocerans are a substantial component (Hessen *et al.*, 1989). Phosphorus contents in treatment T1 favored species with a fast reproduction rate and short life span of organisms, such as Rotifera. Rotifers and small copepods are known to be more tolerant of adverse environmental conditions than the cladocerans (Hannson *et al.*, 2007). As previously, humic acid led to improving water quality and extends algae with essential nutrients which are necessary for their growth; thus, zooplankton growth.

Zooplankton can consume a substantial portion of phytoplankton, but zooplankton- phytoplankton relationships within Wadi-EL Rayan area were variable seasonally and among sites (Nehad and Howayda, 2010). In subtropical and tropical lakes grazing by zooplankton is of little importance for controlling the phytoplankton community structure and biomass (Crisman and Beaver, 1990). The phytoplankton densities in the present fish ponds were correlated with the densities of zooplankton in the same ponds. This may be due to two main factors: the regeneration of the nutrients by zooplankton leads to increase the abundance of phytoplankton (Janik, 1989), and the predation of the fishes on large zooplankton leads to the development of phytoplankton (Elhigzi *et al.*, 1995). Planktivorous fish are known to be size-selective predators that prey selectively on largest zooplankton (Zaret, 1980). The present data are concurrent with this observation since the small rotifers and the nauplius larvae of copepods were the most dominant in fish ponds. Micro zooplankton (rotifers, nauplius larvae, ciliates, and heterotrophic flagellates) develops better when predatory pressure by zooplankton crustaceans is reduced (Richardson *et al.*, 1990).

**Table 4.** Variance and distribution of zooplankton groups (org/L) in earthen ponds stocked with Nile tilapia and subjected to humic acid fertilizer

Zooplankton Groups		T1					T2					T3					T4									
		May	June	July	Aug.	Sept	Oct.	May	June	July	Aug.	Sept	Oct.	May	June	July	Aug.	Sept	Oct.							
Cladocera		3	1	3	2	8	13	—	2	—	—	—	5	8	4	3	2	5	9	11	2	—	—	5	8	13
	%	6.8	1.7	5.3	2.6	10	15	0	5	0	0	0	9.6	14	10.2	5.2	3	6.4	10.8	13	11	0	0	8.4	11.9	20.9
Copepods		7	9	4	16	7	10	2	4	—	11	2	5	9	11	7	18	9	14	1	3	5	10	1	4	
	%	15.9	16	7.1	21	8.7	11	8.6	10	0	20	3.8	8.7	23	19.2	11	23	10.8	16	5.2	9.6	11	17	1.4	6.4	
Ostracoda		3	2	—	2	8	9	—	2	2	—	4	5	—	—	3	5	7	11	—	—	5	3	5	7	
	%	6.8	3.4	0	2.6	10	10	0	5	5.4	0	7.6	8.7	0	0	4.6	6.4	8.4	13	0	0	11	5	7.4	11.3	
Rotifers		24	37	41	55	56	48	18	28	33	38	41	34	22	38	46	41	47	37	14	27	32	41	48	37	
	%	54.5	65	73	71	70	55	78.2	70	89	70	78.8	60	56.4	66.6	71	53	56.6	43	74	87	70	69	71.6	59.6	
Others		7	9	8	2	1	7	3	4	2	5	—	5	4	5	7	9	11	13	2	1	4	—	3	1	
	%	15.9	16	14	2.6	1.2	8	13	10	5.4	9.2	0	8.7	10.2	8.7	11	12	13.2	15	11	3.2	8.6	0	4.4	1.6	
Total number		44	57	56	77	80	87	23	40	37	54	52	57	39	57	65	78	83	86	19	31	46	59	67	62	
	%	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	100	

**Brett and Goldman (1996)** found that removing large and more conspicuous zooplankton, lefts back small crustaceans and small rotifers. **Diana *et.al* (1991)** tested the trophic cascade hypothesis in aquaculture ponds containing tilapia at different densities. They detected that fish predations affect not only the prey, but also lower trophic levels. They found that zooplankton densities, particularly small zooplankton were reduced in ponds without fish.

### Growth Performance

As presented in Table (5), survival rate, final weight, net gain, daily gain and total and net production of fish production were significantly ( $P<0.05$ ) increase in highest stocking density in both with and without humic acid than lowest stocking density with and without humic acid treatments. The survival rate of tilapia was 99.5; 99.5; 100 and 100% for treatment 1, treatment 2; treatment 3 and treatment 4 respectively. The higher survival in humic acid treatments with highest stocking density might result from the availability of nutrient as additional food and shelter for protection from predators.

**Table (5): Growth performance of two stocking density of *O. niloticus* with Humapol (humic acid) fertilizer treatments and without in earthen ponds.**

Treat. items	Treatment 1	Treatment 2	Treatment 3	Treatment 4
Initial weight g	10.0±1.0 <sup>a</sup>	10.0±1.0 <sup>a</sup>	10.0±1.0 <sup>a</sup>	10.0±1.0 <sup>a</sup>
Final weight g	296.4±24.5 <sup>a</sup>	241.1±20.2 <sup>c</sup>	263.7±22.5 <sup>b</sup>	217.6±18.5 <sup>d</sup>
Weight gain g	285.5±23.5 <sup>a</sup>	230.6±20 <sup>c</sup>	253.2±21.2 <sup>b</sup>	207±18 <sup>d</sup>
Daily gain g	1.59±0.15 <sup>a</sup>	1.28±0.12 <sup>c</sup>	1.41±0.12 <sup>b</sup>	1.15±0.1 <sup>c</sup>
Survival rate %	99.5±0.5 <sup>a</sup>	99.5±0.5 <sup>a</sup>	100±0.0 <sup>a</sup>	100±0.0 <sup>a</sup>
Total prod. Pond kg	7416.2±150 <sup>c</sup>	5896.1±140 <sup>d</sup>	13081.4±350 <sup>a</sup>	11644.8±280 <sup>b</sup>
SGR	0.75±0.1 <sup>a</sup>	0.73±0.1 <sup>a</sup>	0.74±0.1 <sup>a</sup>	0.68±0.1 <sup>a</sup>
Feed consumption Kg	11400.7±250 <sup>b</sup>	10976.9±220 <sup>c</sup>	16422.7±300 <sup>a</sup>	16009.2±300 <sup>a</sup>
Feed conversation ratio	1.1±0.1 <sup>c</sup>	1.72±0.1 <sup>b</sup>	1.84±0.1 <sup>b</sup>	2.13±0.1 <sup>a</sup>

Survival, growth, weight gain, daily gain and total production of tilapia were significantly ( $P<0.05$ ) higher in the ponds provided with additional humic acid than those in the same treatments without humic acid Table (5), which may

be attributed to the high productivity and readily available food in the form of humic acid treatments. A similar production enhancement was reported by **(Ramesh *et al.*, 1999)** and **(Azim *et al.*, 2005)** through the provision of additional humic acid. Algae growing on humic acid treatments and the associated bacterial and zooplanktonic biomass can be directly exploited by many herbivorous fish species **(Huchette *et al.*, 2000)**, resulting in a higher fish yield. The present study indicated that humic acid supported fish production, which offers considerable potential for aquaculture production enhancement. The total harvested weights of tilapia were 7416.2; 5896.1; 13081.4; and 11644.8kg/pond for the same sequence treatments Table (5). These results indicated that the humic acid improved water quality, hence led to significantly increase in fish production.

Highest production of different fish species were achieved in the humic acid treatments and had significantly higher ( $P<0.05$ ) values than in the same treatments without substrate without humic acid. Tilapia is regarded as an omnivorous species and capable of feeding on benthic and attached algal and detrital aggregates **(Azim *et al.*, 2003a and b)**.

It is hypothesized that the increases in net yields of fish were largely a result of the availability of easily grazed periphytic food growing on the humic acid. Similar degrees of enhancement of production of *L. rohita* were obtained by **(NFEP, 1997 and Wahab *et al.*, 1999)** through the addition of substrates.

## ***Conclusion***

The present study concluded that effects of humic acid on water parameters have a large advantages since, the physico-chemical parameters studied were improved and the plankton composition documented gave average requirement for all the parameters needed for fish growth and survival, the fluctuation accounted for the different fish yields, while the ponds waters were suitable for fish production.

## ***References***

- Adolf, J. E; Bachvaroff, T. R and Place, A. R. (2009).** Environmental modulation of karlotoxin levels in strains of the cosmopolitan dinoflagellate *Karlodinium veneticum* (Dinophyceae). *J Phycol*, **45**: 176–192.
- APHA. (2000).** Standard Methods for the Examination of Water and Wastewater. 21<sup>st</sup> edition. Eaton, A.D., Clesceri, L.C. and Greenberg, A.E. (eds.). American Public Health Association, Washington, DC.
- APHA. (1985).** American Public Health Association Standard Methods for the examination of water and waste water 15<sup>th</sup> edition. **Pp 476.**



- Avci, M.; Denek, N. and Kaplan, O. (2007).** Effects of Humic acid at different levels on growth performance, Carcass Yields and some biochemical parameters of quails. *Journal of Animal and Veterinary Advances*, **6 (1): 1-4.**
- Azim M.E.; M.A. Wahab; P.K. Biswas; T. Asaeda; T. Fujino; and M.C. Verdegem. 2005.** The effect of periphyton substrate density on production in freshwater polyculture ponds. *Aquac.*, **232: 441–453.**
- Azim M. E.; Verdegem, M. C.; Singh, M.; Dam, A. A. and Beveridge, M. C. (2003b).** The effects of periphyton substrate and fish stocking density on water quality, phytoplankton, periphyton and fish growth. *Aquac. Res.*, **34: 685-695.**
- Azim, M. E.; Verdegem, M. C.; Mantingh, I.; Van Dam, A. A. and Beveridge, M. C. M. (2003a).** Ingestion and utilization of periphyton grown on artificial substrates by Nile tilapia, *Oreochromis niloticus*. L. *Aquac. Res.*, **34: 85–92.**
- Bakhsh, H. K., (2001)** Influence of Humic Acid on Water Quality and Growth of Algae in Freshwater Ecosystem. Masters thesis, Universiti Putra Malaysia.
- Brett, M. T. and Goldman, C. R. (1996).** Ameta-analysis of the freshwater trophic cascade. *Proc. Acad\*Sci.USA*, **93: 7723-7726**
- Carlsson, P.; Graneli, E.; Tester, P. and Boni, L. (1995).** Influences of riverine humic substances on bacteria, protozoa, phytoplankton and copepods in a coastal plankton community. *Mar. Ecol.Prog. Ser.* **127: 213-221**
- Conzonno, V. H. and Cirelli, F. A (1996).** Humic substances and phytoplankton primary production in chascomus pond (Argantina). Facts and Speculations. *Rev. Asoc.Cienc.Nat.Litoral* **27(1): 35-42.**
- Crisman, T. L. and Beaver, J. R. (1990).** Application of planktonic biomanipulation for managing eutrophication in the subtropic. *Hydrobiologia*, **200/201: 177-185.**
- Diana, J. S.; Dettweiler, D. J. and Lin,C.K.(1991).** Effect of Nile tilapia (*Oreochromis niloticus*) on the ecosystem of aquaculture ponds, and its significance to the trophic cascade hypothesis. *Can.J.Fish.Aquat.ScL*,**48:183-190.**
- Elhigzi, F. A. R.; Haider, S. A. and Larsson, P. (1995).**Interactions between Nile tilapia (*Oreochromis niloticus*) and cladocerans in ponds (Khartoum, Sudan).*Hydrobiologia*, **307:263-272.**
- Gutseit, K.; Berglund, O. and Graneli, W. (2007).** Food quality for Daphnia in humic and clear water lakes. *Freshwater Biology*, **52:344–356.**
- Hansson, L. A.; Nicolle, N.; Brodersen, J.; Romare, P.; Nilsson, P. A. and Brönmark, C. (2007).** Consequences of fish predation, migration, and juvenile ontogeny on zooplankton spring dynamics. *Limnology and Oceanography*, **52: 696-706.**

- Hessen, D. O.; Andersen, T.; Lyche, A. (1989).** Differential grazing and resource utilization of zooplankton in a humic lake. *Arch Hydrobiol.* **114:** 321-347.
- Huchette, S. M.; Beveridge, M. C.; Baird, D. J. and Ireland, M. (2000).** The impacts of grazing by tilapias (*Oreochromis niloticus*) on periphyton communities growing on artificial substrate in cages. *Aquac.*, **186:** 45–60.
- Janik, J. J. (1989).** Nutrient recycling in Castle Lake, California: Phytoplankton-zooplankton interactions. Diss. ABST.-TNT.-PT.-B.-and- ENG. **49(8):141.**
- Kankaala, P. (1988).** The relative importance of algae and bacteria as food for *Daphnia longispina* (Cladocera) in a polyhumic lake. *Freshwat. Biol.*, **19:** 285-296.
- Keskitalo, J. and Eloranta, P. (eds.) (1998)** Limnology of Humic Waters. Leiden: Backhuys.
- Klochenko, P.D.; Vasilchuk, T. A.; Medved, V. A. and Vasilenko, O. V. (2011).** Peculiarities of the Influence of Humic Acids on the Development of Plankton Algae. *Hydrobiological Journal*, **47 (1).**
- Meinelt, T. ; Pietrock, M.; Wienke, A. and Volker, F. (2003).** Humic substances and the water calcium content change the toxicity of malachite green. *Journal of Applied Ichthyology*, **19,** 380–382
- NFEP. (1997).** Production enhancement of the Indian major carp, *Labeo rohita* (Ham.) using bamboo trimmings as a substrate for the growth of periphyton. NFEP Paper No. 10. Northwest Fisheries Extension Project, Parbatipur, Dinajpur, Bangladesh.
- Nehad, K. and Howayda, H. A. (2010).** Some Investigations on Zooplankton and Biochemical Contents content of phytoplankton in Wadi El-Rayan Lakes, Egypt. *World Applied Science Journal*, **11(9): 1035- 1046.**
- Prakash, A.; Rashid, M. A. and Jensen, A. (1973).** Influence of humic substances on the growth of marine phytoplankton: diatoms. *Limnol. Oceanogr.* **18:516-524.**
- Prescott, G. W., (1962).** Algae of the Western Great Lakes Area, P: 517. Brown, W. M. C. Com. USA.
- Ramesh, M. R.; Shankar, K. M.; Mohan, C. V. and Varghese, T. J. (1999).** Comparison of three plant substrates for enhancing carp growth through bacterial biofilm. *Aquac. Eng.*, **19:** 119– 131.
- Richardson, W. B.; Wickham, S. A. and Threlkeld, S. T. (1990).** Food web response to the experimental manipulation of a bentivore (*Cypinus carpio*), zooplankton (*Menidia heryllina*) and benthic insects. *Arch. Hydrobiol.*, **119:143-165.**
- Sarir, M. S.; Durrani, M. I. and Mian, A. I. (2006).** Effect of the source and rate of humic acid on phosphorus transformations. *Journal of Agricultural and Biological Science*, **1:** 29-31

- Sánchez-Marín, P. and Beiras, R (2011).** Adsorption of different types of dissolved organic matter to marine phytoplankton and implications for phytoplankton growth and Pb bioavailability. *Journal of Plankton Research*, **33**: 9Pp. 1396-1409
- Shaker, I. M. (2008).** Effect of using different types of organic manure (compost; chicken, mycelium) and mineral fertilizer on water quality, plankton abundance and on growth performance of *Oreochromis niloticus* in earthen ponds. Abbassa. *Int. J. Aqua.* (1A): **203-227**.
- Shaker, I. M. and Mahmoud, A. A. (2007).** The biological load of silver carp cages in Nile River and effects of their on water quality & growth performance. *Egypt. J. Aquat. Biol. & Fish.*, **11 (2)**: 119-143.
- Shaker, I. M. and Abdel-Aal, M. (2006).** Growth performance of fish reared under different densities in semi-intensive and extensive earthen ponds. *Egypt. J. Aquat. Biol. & Fish.*, **10 (4)**: 109-127.
- Shaker, I. M.; Mona, A. H. and Mahmoud, A. A. (2009).** Effect of periphyton substrate on water quality and growth performance of Nile tilapia in earthen ponds. Abbassa Int.J. Aqua., Special Issue for Global Fisheries & Aquaculture Research Conference, Cairo International Convention Center, 24–26 October, pp **741-763**.
- Shaker, I. M; Abou Zeid, M. Y. and Batran, A. (2013):** Effect of using periphyton substrate (bamboo stems) on water quality, phytoplankton, zooplankton, periphyton and growth performance for tilapia, mullet and catfish in earthen ponds. Abbassa, *Int., J. Aqua.*, **6 (No 1)**: 108-139.
- Shannon, C. R. and Weaver, W. (1949).** The Mathematical Theory of Communication. Urbana University Press, Urbana, Illinois, 117 pp.
- Singh, C. P. and Amberger, A. (1991).** Solubilization and availability of P during decomposition of rock phosphate enriched straw and urine. *Biol. Agric. Hort*, **7**: 261.
- Smith, D. W. and Pledrahita, R. H. (1988).** The relation between phytoplankton and dissolved oxygen in fish ponds. *Aquaculture*, **68**: 249-265.
- Statistical Analysis System (SAS). (2000).** SAS program Ver 6.12, SAS institute incorporation, Cary, NC 27513 USA.
- Steinberg, T. M; Timofeyev, M. A; Bittner, .M. and Menzel, R. (2008)** Review Series Humic Substances Part 2: Interactions with Organisms. *Env Sci Pollut Res*, **15 (2)**: 128 – 135.
- Sun, B. K; Tanji, T. and Unno, H. (2006).** Extinction of cells of cyanobacterium *Anabaena circinalis* in the presence of humic acid under light. *Appl Microbiol Biotechnol.*, **72**: 823-828.
- Taniguchi, G. M.; Bicudo, D. C. and Senna, P. A. C. (2005).** Gradiente litorâneo-limnético do fitoplâncton e ficoperifíton em uma lagoa da planície de inundação do Rio MogiGuaçu. *Revista Brasileira de Botânica.*, **28**: 137-147.

- Thompson, F. B.; Abreu, P. C. and Wasielesky, W. (2002).** Importance of biofilm for water quality and nourishment in intensive shrimp culture, *Aquac.*, **203**: 263– 278.
- Vrana, D. and Votruba, J. (1995).** Influence of soluble humic substances on the of Algae and Blue-Green Algae. *Folia Microbiol.*, **40 (2)** : 207-208.
- Wahab, M. A.; Mannan, M. A.; Huda, M. A.; Azim, M. E.; Beveridge, M.C. and Tollervey, A.G. (1999).** Effects of periphyton grown on bamboo substrates on growth and production of Indian Major carp rohu (*Labeo rohita* Ham.). *Bangladesh J. of Fisheries Res.*, **3**: 1-10.
- Wet, L .F. and Visagie, W. (2010).** Evaluating CHD-FA Carbohydrate-Derived Fulvic Acid for Use in Diets of Mozambique Tilapia *Oreochromis Mossambicus*. Feed Technology Group Division Aquaculture University of Stellenbosh South Africa.
- Zaret,T. M. (1980).** Predation and freshwater communities. New Haven and London,Yale Univ. Press,**187**pp.

## تأثير استخدام حامض الهيوميك على الهائمات وجودة المياه ونمو أسماك البلطي النيلي في أحواض الإستزراع السمكى

محمد جمعه بطاح<sup>1</sup>، عايدة محمد عبدالله ضوة<sup>2</sup>، أيمن أنور عمار<sup>2</sup>، محمد مختار سلامة<sup>2</sup>

1- قسم النبات شعبه الميكروبيولوجى كلية العلوم جامعة بنها.

2- المعمل المركزى لبحوث الثروة السمكية بالعباسة.

تم إجراء هذه الدراسة في وادى الرايان، محافظة الفيوم، جمهورية مصر العربية بغرض تقييم تأثير حمض الهيوميك على نمو الهائمات، وجودة المياه، ومظاهر النمو والإنتاجية الكلية لأسماك البلطي النيلي. وقد تمت الدراسة من خلال استخدام ثمانية أحواض ترابية تقدر مساحة كلا منها اثنين فدان لمدة 24 أسبوع. تم تقسيم هذه الأحواض السمكية إلى أربعة معاملات، (حوضين لكل معاملة). تم استزراع الاسماك فى المعاملتين (الأولى والثانية) بزريعة البلطى بمعدل 14000 زريعة/ فدان أما فى المعاملتين ( الثالثة والرابعة) فقد تم الإستزراع بمعدل 28000/ فدان. المعاملات الثانية والرابعة كانت خالية تماما من أى تسميد غير أنها تغذى بعلف تجارى يحتوى على نسبة بروتين قدرها 27% بروتين بمتوسط يومى 3 % من وزن الجسم مرتين يوميا. بينما تم تسميد المعاملات الأولى والثالثة فقط بحامض الهيوميك بمعدل 3 كجم / فدان أثناء إعداد الأحواض بالإضافة الى 1.5 كجم من سماد الهيوميك لكل طن عليقة أثناء التغذية.

وقد تم تلخيص النتائج كالتالى:

- 1- لا توجد فروق ذات دلالة إحصائية في درجة حرارة الماء أو القلوية الكلية أو عسر الماء أو درجة الملوحة.
- 2- تركيزات الأمونيا ، والأوكسجين الذائب، الفوسفات الذائب، الكلوروفيل أعلى فى كل من المعاملتين الأولى والثالثة عنه فى المعاملتين الثانية والرابعة.
- 3- تركيزات النترات والنترت أعلى بأحواض المعاملتين الرابعة والثانية عنه فى المعاملتين الأولى والثالثة .
- 4- سجلت أقل قيمة للشفافية فى كلا من المعاملة الاولى والثالثة بينما رصدت أعلى قيمة لها بالمعاملة الرابعة.
- 5- أعلى تركيز من الهائمات ( النباتية والحيوانية ) كان بالمعاملة الأولى ثم الثانية يليه المعاملة الثالثة.
- 6- كان أعلى وزن نهائى كان بالمعاملة الأولى والثالثة بينما كان الأقل فى المعاملة الرابعة.