NUTRITIONAL CONTENT OF SELECTED MACROALGAE OF THE SOUTH-WEST COAST OF INDIA

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Abstract:

During the present work, thirty-one species of macroalgae collected from different locations on the south-west coast of India were subjected to nutritional analysis to determine total crude protein, total crude lipids, total soluble sugars, starch, total free amino acids, iodine content, and vitamin C content. It could be derived that the protein content varied from a minimum of 10.8±0.7 mg g-1 in the red alga Amphiroa fragilissima to a maximum of 180.0±0.7 mg g⁻¹ in Chaetomorpha linum (Chlorophyceae), followed by118.0±0.3 mg g⁻¹ in Sargassum marginatum (Phaeophyceae). The lipid content was recorded to be a maximum of 150.0±0.2 mg g⁻¹ in the brown alga Padina tetrastroematica and a minimum of 8.0±0.4 mg g⁻¹ in the calcareous red alga Cheilosporum spectabilis. The maximum free amino acid concentration was found to be 17.06 ± 0.6 mg g⁻¹ in the green alga Caulerpa sertularioides, the maximum value for Vitamin C content was 11.0±0.4 mg 100g⁻¹ in the green alga Caulerpa chemnitzea, and the maximum iodine content of 86.90±1.0 mg 100g⁻¹ in green alga Caulerpa peltata. The Vitamin C content was found to be reasonably high in almost all algal species studied, as also the iodine content. The overall observation shows that marine macroalgae are rich sources of Proteins, Carbohydrates, Amino acids, Vitamin C, and Iodine, hence can have great implications as source materials in the preparation of food, food supplements, and nutraceuticals.

Keywords: Marine macroalgae, South-west Indian coast, Nutritional composition

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Introduction

Seaweeds form a very important renewable resource in the marine environment and have been a part of human civilization from time immemorial. The reports on the uses of seaweeds have been cited as early as 2,500 years ago in Chinese literature (Tseng, 2004). The history of seaweed utilization for a variety of purposes is indicative that some of their constituents are superior and valuable in comparison to their terrestrial counterparts. Seaweeds synthesize an array of chemicals, some of which are the only natural resources to produce useful substances such as agar-agar, carrageenan, and alginates. Seaweeds form a valuable source of food containing proteins, carbohydrates, lipids, iodine, vitamins, and bioactive molecules with potential as nutritional supplements, pharmaceuticals, cosmeceuticals, fine chemicals, and enzymes (Pereira, 2018a, b; Peñalver et. al. 2020). Marine macroalgae are also rich sources of several bioactive substances like polysaccharides, polyphenols, phytochemicals, and polyunsaturated fatty acids with potential therapeutic uses against inflammation, cancer, oxidative stress, allergies, diabetes, thrombosis, obesity, hypertension, lipidemia, and many degenerative diseases (Tanna and Mishra, 2019); hence these marine plants help to improve the quality of life (Holdt & Kraan, 2011; Shannon & Abu-Ghannam, 2019). Due to their various health benefits, there is an increasing interest in utilizing them as culinary ingredients. Parallelly the everincreasing population is compelling humans to opt for non-conventional and alternative food resources such as seaweeds. The world's annual production of edible seaweeds through aquaculture is 6 million tons of fresh weight (Fleurence, 1999). In all 152 seaweed species have been utilized for food purposes globally, of which 83 are from Rhodophyta, 40 from Phaeophyta, and 29 from Chlorophyta. The main edible seaweeds are Japanese kelp (Laminaria japonica),

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Kombu (Sacharinna spp.), Gracilaria spp., Nori Nei (Porphyra spp.), Eucheuma seaweeds (Eucheuma spp.), Laver or Nori (Porphyra tenera), Wakame (Undaria pinnatifida), Elkhorn Sea moss (Kappaphycus alvarezii), Hiziki (Sargassum fusiforme), Umudggasari (Gelidium amansii), and Gamtae (Ecklonia cava), among others (Sarker et al., 2021; Zhang et. al., 2022).

India (08.04 - 37.06 N and 68.07 - 97.25 E), a tropical South Asian country has a stretch of about 7500 km coastline, excluding its island territories with 2 million km² Exclusive Economic Zone (EEZ) and nine maritime states. The seaweed flora of India is highly diversified and comprises mostly tropical species, but boreal, temperate, and subtropical elements have also been reported. A total of 830 species of marine algae have been recorded from different parts of the Indian coast including the Andaman-Nicobar and Lakshadweep Islands (Reddy et al., 2014). Many of the rocky beaches, mudflats, estuaries, coral reefs, and lagoons along the Indian coast provide ideal habitats for the growth of seaweeds. In India, though seaweeds have not gained much popularity as human food there are stray reports of their use in food formulations. In some coastal places of Tamil Nadu, (Tiruchandur- Kanyakumari sector), southern India, people are reported to take "Seaweed Ganji" (water in which seaweed Hypnea sp. is boiled) for getting rid of stomach disorders, as well as seaweed extracts from species of Ulva in the preparation of sweets. In Madurai city of Tamil Nādu, Agar is added to a milkshake-like summer drink called 'Jigarthanda' (in place of gelatine) to give it its thick consistency, along with sabja seeds (sweet basil), sarsaparilla (Smilax sp.) or rose syrup, milk, and ice cream (Cordelia, 2021). By keeping in mind that seaweeds form a potential alternative vegetarian diet in a developing country like India, it is worthwhile to review in detail the nutritional value of seaweeds as human food in the new millennium. Since the author has been working extensively on the marine algal resources of Kerala, the present

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work was undertaken to analyse the nutritional composition of selected marine macroalgae of the southwest coast of India, predominantly the Kerala coast.

Materials and Methods

Sample collection

Kerala state is situated on the southwest coast of India between Lat. 8020' to 12°51'N and Long. 74°53'to 77°33'E; the length of the coastline is 560 km stretching from Thiruvananthapuram in the south to Kasaragod in the north. The substratum is mostly rocky in nature which facilitates rich and varied algal growth. The bit of coastline extending from Cape Comorin in the peninsular tip to Thiruvananthapuram also supports rich and diverse algal growth. Using a 0.5m² wooden quadrate, samples of the algae from the selected/productive areas on the southwest coast of India (Cape Comorin in the peninsular tip to Kasaragod in the northern tip of Kerala) were collected (Fig.1) and different species were noted down separately and their biomass studied (August 2018- July 2019). The identification of the algal samples was carried out with the help of standard flora (Iyengar, 1927; Boergesen, 1935; Boergesen, 1938; Krishnamurthy, 1971; Desikachary, 1967; Untawale et al., 1983; Nair et al., 1986; Kaliaperumal, 2006). Based on the above survey, 31 species were later selected for biochemical analyses. The voucher specimens were given accession numbers and deposited at the 'Marine Algae Herbarium,' Dr. KSM Centre for Algal Biotechnology, Thiruvananthapuram.

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Hydrographic studies

At the time of algal sampling, atmospheric, and surface water temperatures were recorded using standard thermometers. Hydrographic parameters such as salinity, pH, and dissolved oxygen (**Martin**, 1969), and nutrients such as phosphates, nitrates, nitrites, and silicates (**Strickland and Parson**, 1972) were also estimated. Light penetration was measured using a Secchi disc. For the convenience of the study, the year is divided into three seasons viz. pre-monsoon (February to May) the monsoon (June to September), and the post-monsoon (October to January) seasons.

Chemical analysis

Thirty-one species of edible marine algae (Green, Brown, and Red) collected from the study area were subjected to biochemical analyses. The collected thalli were washed thoroughly with fresh water 3 to 4 times to remove associated fauna, epiphytes, and debris and dried under shade. These were then powdered and stored in airtight plastic containers (food grade) until the time of analysis. Total crude protein was estimated using Folin - Phenol reagent with bovine serum albumin serving as standard (Lowry, 1951); total lipid content was determined by extraction with the chloroform-methanol mixture using a separating funnel (Bligh and Dyer, 1959); total soluble sugars (pentoses, hexoses, disaccharides including sucrose, maltose, lactose, and hexuronic acid) by using Anthrone and Phenol-sulphuric acid reagents (Yemm & Willis, 1954); and total free amino acids was estimated by modified Ninhydrin method byYemm and Cocking, 1955 (Hyman, 1957), and starch converted to sugar by extraction with Perchloric acid and calculated in terms of glucose equivalent; a conversion factor is used to convert glucose to starch (Mahadevan and Sridhar, 1996). The iodine content in the seaweeds was estimated by iodometric titration using Sodium

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thiosulphate (UNICEF *et al.*, 1995). Vitamin C was determined by use of an oxidation-reduction reaction by redox titration using potassium iodate in the presence of Potassium iodide. (Tee *et al.*, 1988).

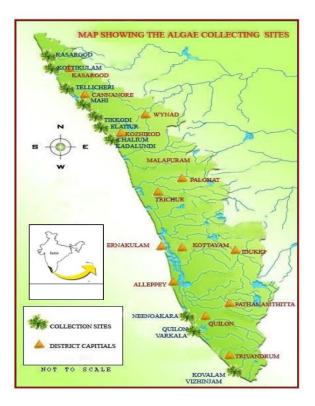


Fig. 1: Algae collection sites of SW Coast of India

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Statistical Analysis

For biochemical estimation, triplicate samples of each species were analysed, the results of which were calculated as mean \pm Standard Error (SE).

Results and Discussion

Studies on the seasonal variation in various physicochemical parameters such as salinity, pH, Dissolved Oxygen, and Nutrients like Phosphate, Nitrate, Nitrite, and silicate at the collection sites were carried out coinciding with the algal collection during the period from August 2018 to July 2019

Temperature: - Distinct seasonal fluctuations were noted in the distribution of both atmospheric and surface water temperatures. Atmospheric temperature varied from 26° C to 31.5° C and surface water temperature from 27.5° C to 29° C along the algae collection locations during the period of study.

Light penetration: - The Sechi Disc transparency was found to range from 190 to 380 cm during the period of study.

Salinity: - The salinity values varied from 27.25%⁰ to 34.25%⁰ along the area studied. High values were generally recorded during the pre-monsoon season. Low values were recorded during the monsoon periods

pH: - the pH varied from 6.53 to 8.04 along the coast. Water was found to be alkaline at the Varkala and Thirumullavaram coasts.

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Phosphate values were moderate, found to vary from 0.97 to 4.99 μ g.at. L⁻¹. The higher phosphate values were recorded during periods of heavy rainfall. This may be due to the added nutrients through drainage and upwelling.

Nitrate level showed a wide variation from 3 to 36 μ g.at. L⁻¹ The highest nitrate concentrations were recorded corresponding to heavy rainfall. The monsoon rains and the resultant drainage could be a contributing factor in increasing the nitrate concentration during this period.

Nitrite varied from 0.32 to 17.0 μ g.at. L^{-1.} The nitrite values were always found to be much lesser than the nitrate values and higher values were recorded during the monsoon seasons.

Silicate content generally read very high, especially during the monsoon periods; it was found to vary from 47.47 to 321 μ g.at. L⁻¹. It is now well known that the freshwater is rich in Silica than seawater and freshwater discharge during monsoon season has been accepted as a source of silicate during monsoon season.

During the present work, it could be observed that, though the heavy monsoon rains coupled with a turbulent sea proved to be unfavourable for the growth of many of the edible algal species, brown algae like *Chnoospora bicanaliculata* and *Sargassum* spp.; green algae like *Ulva* spp., *Enteromorpha prolifera*, *Chaetomorpha antennina & Valoniopsis pachynema*, and red algae like *Gracilaria corticata* were found to predominate the coast to the exclusion of other algae during the monsoon period.

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Studies correlating the physicochemical properties and macroalgal abundance of the Kerala coast have been meager (Nair *et al.*, 1986, Chennubhotla, 1991; Maya, 1990; Maya 2010; Shynu *et al.*, 2012; Sushanth & Madaiah, 2014; Aravindh & Ruban, 2019). Since the present paper primarily deals with the nutritional composition of seaweeds, in-depth environmental details have not been discussed.

Nutritional composition

Among the 31 species of marine algal species chosen for nutritional analyses Caulerpa peltata, C. sertularioides, Chaetomorpha linum, C. antennina, Enteromorpha prolifera, Padina tetrastromatica, sargassum spp., Gracilaria spp., Acanthophora spp., Champia parvula, and Rhodymenia sp. are the most edible species found worldwide. The seaweed species were chosen for analyses based on edibility, standing crop, and seasonal availability. Table 1. furnishes the list of 31 algae analysed and the results of the nutritional analysis of seaweed species collected from the southwest coast of India. The protein content of the algae varied from 17.1±1.0 to 180.0±0.7 mg g-1 in Chlorophyceae; 49.3±0.2 to 118.0±0.4 mg g⁻¹ in Phaeophyceae and from 10.8±0.7 to 90.3±0.4 mg g⁻¹in Rhodophyceae. The protein content of seaweeds is found to differ according to the species. Generally, the protein fraction of brown seaweeds is low (3 to 15% dry wt.) as compared with that of green seaweeds (10 to 47% dry wt.) (Arasaki and Arasaki, 1983). In some green seaweeds like Ulva, the protein content varies between 10 to 26% of the plant (dry wt.) (Fujiwara-Arasaki et al., 1984). During the present work, Ulva fasciata which is one of the commonest green seaweeds of the Indian coast is found to have a protein content of about 9% (dry wt.) and hence its suitability was evaluated in the preparation of protein-rich biscuits and other food products during this work.

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SL	Name of algae	Protein	Lipid	Carbohydrate	Starch	Amino	Vitamin C	Iodine
		mg. g ⁻¹	mg. g ⁻¹	(Total	mg. g ^{.1}	acid mg. g	mg. 100g-1	mg. 100g-1
No				soluble)		1		
				mg. g ⁻¹				
	CHLOROPHYTA	Mean±SE (n=3)						
1	Caulerpa peltata	77.0±3.0	14.0±0.8	3.8±0.4	4.0±0.4	5.9±0.2	8.5±0.2	86.9±1.0
2	Caulerpa sertularioides	64.3±3.0	73.4±0.6	31.1±0.8	28.0±0.4	17.1±0.6	3.4±0.1	52.9±1.0
3	Caulerpa chemnitzea	124.0±1.0	32.0±1.0	19.6±0.8	18.0±0.5	9.3±0.3	11±0.4	61.3±1.0
4	Chastomorpha antennina	58.6±2.0	11.0±1.0	13.8±1.0	12.9±1	3.5±0.3	4.0±0.1	47.2±1.0
5	Chastomorphalinum	180±0.7	113.0±0.8	10.6±0.9	9.0±0.8	1.6±0.1	3.0±0.1	50.8±2.0
6	Cladophora sp.	46±0.1	4.0±0.4	5.8±1.0	5.0±0.8	10.6±0.5	8.0±0.0	86.7±1.0
7	Ulva fasciata	91.0±0.1	28.0±1	34.1±1.0	36.1±0.9	2.5±0.1	4.2±0.1	27.5±1.0
8	Enteromorphaprolifera	17.1±1.0	20.0±1.0	6.9±0.4	5.4±0.4	4.8±0.3	7.5±0.1	50.8±0.1
9	Valoniopsis pachynema	43.7±0.7	20.0±04	38.4±1.0	34.0±0.9	2.2±0.2	7.3±0.5	36.0±1.0
	PHEOPHYTA							
10	Padina tetrastromatica	51.7±1.0	150.0±2.0	12.0±1.0	10.3±1.0	2.6±0.3	7.3±0.1	25.5±2.0
11	Chnoospora	49.3±2.0	12.6±.08	16.6±1.0	14.9±1.0	6.0±0.3	6.1±0.1	33.8±1.0
	bicanaliculata							
12	Sargassum marginatum	118.0±0.3	12.0±0.6	40.8±1.0	36.3±0.7	3.8±0.3	8.6±0.1	23.3±4.0
13	Sargassumtenerrimum	87.0±2.0	9.0±0.8	24.0±0.8	21.0±0.5	3.7±.4	9.6±0.1	19.1±3.0
14	Sargassum cinereum	117.0±0.4	12.0±1.0	20.6±0.2	19.3±0.8	5.4±0.3	5.3±0.1	23.2±1.0
15	Sargassumwightii	65.7±0.5	14.0±0.4	12.5±0.2	10.7±0.2	7.06±0.2	6.4±0.2	36.0±1.0
	RHODOPHYTA							
16	Spyridia insignis	29.3±1.0	45.0±1.0	9.0±0.4	8.0±0.9	7.6±0.2	5.2±0.1	36.0±.01
17	Spyridia hypnoides	24.0±0.4	34.0±0.8	39.6±0.8	35.7±0.8	13.2±0.4	10.8±0.1	33.8±1.0
18	Centroceras clavulatum	31.7±0.3	70.0±2.0	21.5±0.9	19.2±0.9	15.6±0.6	5.4±0.1	35.9±1.0
19	Amphiroa fragilissima	10.8±0.7	10.0±2.0	4.3±0.8	3.1±0.7	3.2±0.1	1.4±0.1	25.5±3.0
20	Gelidiopsis variabilis	34±0.2	93.4±1.0	27.1±1.0	27.0±0.5	35.6±0.3	3.0±0.1	33.8±1.0
21	Grateloupialithophila	40.7±0.2	40.0±0.8	37.8±0.2	35.3±0.5	4.4±0.3	3.7±0.1	49.0±1.0
22	Laurencia brongniartii	56.3±1.0	80.0±2.0	17.9±0.5	15.8±0.1	4.9±0.2	5.0±0.1	34.0±3.0
23	Accanthophora specifera	29.3±3	80.0±2.0	24.9±1.0	21.9±1	8.1±0.5	3.7±0.1	52.9±1.0
24	Acanthophora	26.0±0.2	NA	24.4±1.0	NA	NA	NA	NA
	najadiformis							
25	Rhodymeniasp.	52.0±0.2	20.0±1.0	15.2±0.3	14.3±0.8	9.8±0.4	4.6±0.1	33.8±1.0
26	Champiaparvula	68.33±3	60.0±1.0	27.4±1.0	23.3±1	8.1±0.3	2.4±0.1	52.9±1.0
27	Gracilaria corticata	22.1±1.0	142.0±2.0	8.5±0.9	6.3±0.8	1.7±0.2	4.4±0.1	38.1±0.0
28	Gracilaria fergusonii	55.66±10	100.0±2.0	39.7±0.5	36.3±0.5	4.8±0.1	4.0±0.1	27.6±1.0
29	Gracilaria foliifera	56.3±0.9	40.0±1.0	11.4±0.9	9.6±0.8	10.2±0.3	3.3±0.1	40.2±1.0
30	Chelosprumspectabilis	31.33±10	8.0±0.4	2.20±1.0	2.0±0.1	2.2±0.1	2.1±0.1	23.26±1.0
31	Enantiocladiasp.	90.33±0.4	20.0±0.8	22.4±1.0	20.8±0.8		6.8±0.1	NA

Table 1: Nutritional content of selected marine algae of the south-west coast of India

NA- Not analysed

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According to Paul et al. (2007), the protein levels in seaweeds vary according to the season and the species. For most species, aspartic and glutamic acids constitute a large part of the amino acid makeup of these proteins. These levels are highest in brown seaweeds, with red seaweeds having the lowest During the present inquiry, red seaweeds were found to have a amounts. comparatively low protein content as opposed to the study by Dere et al. (2003) where brown algae were found to have low protein content. High protein content in Rhodophyta, moderate in Chlorophyta, and lowest in Phaeophyta was recorded in seaweeds of the Tamil Nadu coast by Rameshkumar (2013). During the present analysis, the protein content in the seaweeds was found to be comparatively lower than that recorded for the Kerala coast by Nair et al. (1991). The protein content in Ulva fasciata was lower compared to earlier studies along Indian coasts (Sitaker Rao and Tipnis, 1964; Dhargalkar et al., 1980; Nair et al., 1991). Similarly, the protein value for Acanthophora specifera is much lower than that reported by Rameshkumar (2013). During the present analysis, the highest protein content of 18% was recorded in Chaetomorpha linum, followed by Sargassum marginatum (11.8%) and S. cinereum (11.7%). The least protein content (1% & 3%) was found to be in the calcareous red algae Amphiroa fragilissima and Cheilosporum spectabilis respectively. Green and red seaweeds have higher protein contents than brown seaweeds, as high as 47% of their dry weight (Černá 2011; O'Connor et al., 2020). Porphyra spp., Pyropia spp., Palmaria palmata, and Ulva spp. are the seaweeds richest in protein (Pereira 2011; Taboada et al. 2013; Angell et al., 2016). According to Ganesan et al. (2019) brown seaweeds contain the least protein while moderate concentrations of protein are reported from green seaweeds, and the highest content is estimated in red seaweeds.

Overall, lipids make up about 1-5% of seaweed dry weight, and Polyunsaturated fatty acids (PUFA) constitute a significant part of the seaweed

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lipid profile (Penalver et al., 2020). During the present analysis, the lipid concentration was found to be low in most of the algal species studied varying from 1.0 to 15%, which is in tune with the findings of Nehal et al. (2011). The values were found to vary from a minimum of 4.0 ± 0.4 mg g⁻¹ to a maximum of 113.0±0.8 mg g⁻¹ in Chlorophyceae; from 9.0±0.8 to 150.0±2.0 mg g⁻¹ in Phaeophyceae and from 8.0±0.4 to 142.0±2.0 mg g⁻¹ in Rhodophyceae. Present lipid values are much higher than that given by Nair et al. (1991) for most of the algal species of the Kerala coast, though the lipid value for Caulerpa peltata (1.4%) is in agreement with the above work. The lipid values for *Chaetomorpha* antenna (1.1%) and Valoniopsis pachynema (2%) reported at present are much higher than those given by Nair et al. (1991). In the nutritional studies carried out on the algal species of the Bay of Bengal coast (Rameshkumar, 2013), Caulerpa racemose was found to have a lipid content of 19.1%, Ulva fasciata - 10.5%, Chnoospora minima - 0.9%, Padina gymnospora- 11.4% and Acanthophora specifera 2.1%. During the present analysis, Caulerpa spp. were found to have lipid contents ranging from 1.4 to 7.3%, and U. fasciata with a value of 2.8%, which is lower than that in the former report. Nevertheless, the values for Chnoospora bicanaliculata (1.2%), Padina tetrastromatica (15%), and Acanthophora specifera (8%) during the present work are higher than that reported by Rameshkumar (2013). The lipid content reported for Gracilaria foliifera, Sargassum wightii, S. tenerrimum, and Ulva lactuca by Manivannan (2008 a, b) is like the results of the present analysis. In general, seaweeds exhibit low lipid contents (Dawes, 1998), and the differences in the lipid content could be attributed to factors like climate, the geography of development to methods of extraction (Oritz et al., 2006). Figs. 2 to 4 furnish the protein and lipid contents in different algal classes. Overall, lipids make up about 1-5% of seaweed dry weight (Penalver et al., 2020). A recent study showed that the lipid content of red alga

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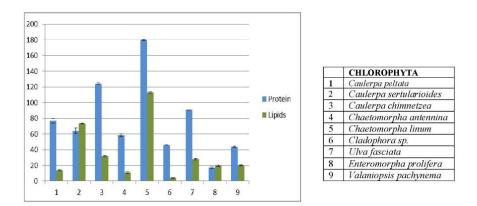
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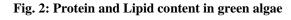
Asparagopsis taxiformis was 2.9–6.2 g per 100 g dry wt., which contributed about 9.5% to the RDI (Mellouk *et al.*, 2017; Nunes *et al.*, 2019).

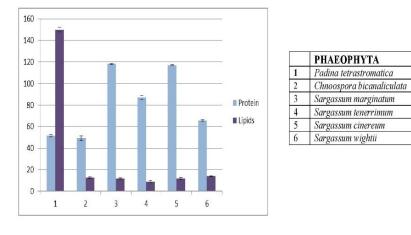
The carbohydrates in seaweeds consist of monosaccharides, disaccharides, and polysaccharides. The first two are simple sugars with low molecular weight, soluble in water such as glucose, galactose, fucose, ribose, and mannitol. Polysaccharides are complex sugars with high molecular weight insoluble in water and these include cellulose, starch, agar, alginate, and carrageenan. The total polysaccharide content in macroalgae ranges from 4% to 76% of dry weight (Holdt and Kraan, 2011). The highest carbohydrate contents are found in red algal genera like Porphyra and green seaweed genera such as Ulva (Holdt and Kraan, 2011), these polysaccharides represent the main nutritive storage in algae with numerous applications. Thus, the polysaccharide profile in seaweeds is quite extensive and deserves special study by itself. Seaweed polysaccharides have numerous beneficial properties such as probiotic activity, inhibition of viruses, suppression of gastrointestinal inflammation, anticancer properties, reduction in cholesterol uptake, and anti-glycosidase activity (Rajapakse & Kim, 2011; Wang et al., 2012; Necas & Bartosikova, 2013; Cotas et al., 2020; Daub et al., 2020). In addition, seaweed fibres contain negligible amounts of starchy carbohydrates, resulting in a lower glycaemic load, which is beneficial in regulating the glycaemic index of diabetic patients (Wee & Henry, 2020).

The total soluble carbohydrates during the present analysis were found to vary from a minimum of 3.80 ± 0.4 mg g⁻¹ in *Caulerpa peltata* to a maximum of 38.40 mg g⁻¹ in *Valoniopsis pachynema* (Chlorophyceae); from 12.0 ± 1.0 mg g⁻¹ in *Padina tetrastromatica* to $40.8\pm$ mg g⁻¹ in *Sargassum marginatum* (Phaeophyceae) and from 2.2 ± 1.0 mg g⁻¹ in *Cheilosporum spectabilis* to 39.6 ± 0.8 mg g^{-1 in} *Spyridia hypnoides* (Rhodophyceae).

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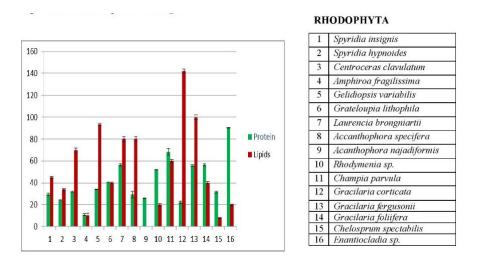


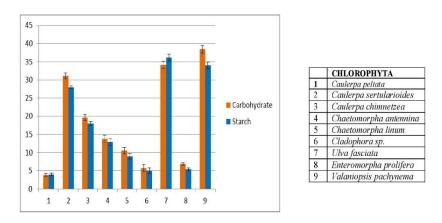
Fig. 3: Protein and Lipids in red algae

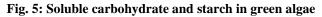
The values for soluble carbohydrates were found to be much lower than the total carbohydrate values reported by **Nehal** *et al.* (2011); there appears to be not much variation in carbohydrate content among the different algal groups. According to the report given by **Nehal** *et al.* (2011), the membranous Phaeophycean species are found to have lower carbohydrate content than the Chlorophycean members. Studies by **Marinho-Soriano** *et al.* (2006) showed that changes in carbohydrate content can be observed over a period. **Rosenberg and Ramus** (1982), related carbohydrate synthesis to periods of maximum growth, increased photosynthetic activity, and reduction in protein contents, suggesting a link between seaweed growth and carbohydrate content. During the present study, the variation in starch content was like that in soluble sugars. It was found to vary Egyptian J. of Phycol. Vol. 24, 2023 - 175 - from 4.0 ± 0.4 mg g-1 in *Caulerpa peltata* to 36.1 ± 0.9 mg g-1 in *Ulva fasciata*; from 10.3 ± 0.2 mg g-1 in *Padina tetrastromatica* to 36.3 ± 0.7 mg g-1 Sargassum *marginatum* (Phaeophyceae), and from 2.0 ± 0.1 mg g-1 in *Cheilosporum spectabilis* to 36.3 ± 0.5 mg g-1 in *Gracilaria fergusonii* (Rhodophyceae). Figs. 5 to 7 furnish the soluble sugars and starch content in different classes of seaweeds. According to the work by **Meghanath** *et al.* (**2019**), on green macroalga *Ulva ohnoi*, the starch content was found to vary from 1.59% to 21.4% depending on the growth conditions and seasons. Besides, nutrient starvation was also found to significantly increase the starch content. Different seaweed groups contain a wide group of polysaccharides whose chemical composition and amount vary within various seaweed species (**Misurcova, 2011**). Hence deciphering carbohydrate content in seaweeds, by itself is an extensive topic to study, based on seasonal studies, geographic locations, growth stages of the plants, and methods of analyses.

Essential amino acids help to build up muscles, support their functioning, and regulate the blood sugar level (**Breitman** *et al.*, **2011; Hayashi** *et al.*, **2018**). Essential amino acids such as leucine, valine, and threonine are abundant in red seaweed species such as *Porphyra dioica*, *Porphyra umbilicalis*, and *Gracilaria vermiculophylla* (**Machado** *et al.*, **2020**). The total free amino acid content was found to vary from a minimum of $1.6 \pm 0.1 \text{mg g}^{-1}$ in *Chaetomorpha linum* to a maximum of $17.6\pm0.6 \text{ mg g}^{-1}$ in *Caulerpa sertularioides* (Chlorophyceae); from 2.6 ± 0.3 in *Padina tetrastromatica* to $7.06\pm0.2 \text{mg g}^{-1}$ in *Sargassum wightii* (Phaeophyceae) and from $1.7\pm0.2 \text{ mg g}^{-1}$ in *Gracilaria corticata* to $35.6\pm0.3 \text{ mg}$ g⁻¹ in *Gelidiopsis variabilis* (Rhodophyceae). The amino acid values obtained are found to be far less than that obtained by **Dinesh** *et al.* (**2007**). Free amino acids have been found important contributors to the taste of seaweeds. They impart a sweet, sour, or bitter taste depending on the amino acid and its concentration (**Kawaai** *et al.*, **2012**).

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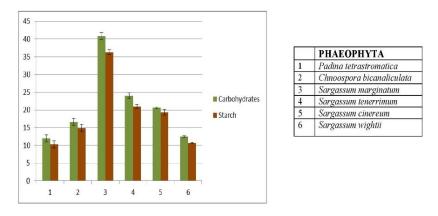


Fig. 6: Soluble carbohydrate and starch in brown algaeEgyptian J. of Phycol. Vol. 24, 2023- 177 -

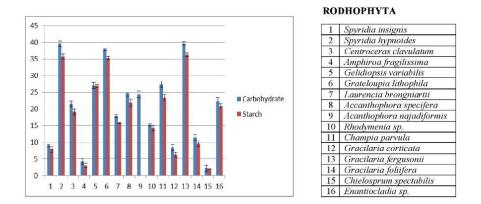


Fig. 7: Soluble carbohydrate and starch in red algae

During the present work, the Vitamin C content was found to be reasonably high in almost all algal species analysed. It was found to vary (in mg100g ⁻¹dry wt.) from the least value of 3.0±0.1 in Chaetomorpha linum to a maximum value of 8.5±0.2 in Caulerpa peltata (Chlorophyceae); from 6.1±0.1 in Chnoospora bicanaliculata to 9.6±0.1 in Sargassum tenerrimum (Phaeophyceae) and from 1.4±0.1 in Amphiroa fragilissima to 6.8±0.1 in Enantiocladia sp. (Rhodophyceae). However, the values are found to be much lower than that recorded for the marine algal species of the Gulf of Mannar by Dinesh et al. (2007) but comparable to that obtained for the benthic algae of Sundarbans (Chakraborthy & Santra, 2008). Skrovankova (2011) suggests that seaweeds growing closer to the surface level contain higher levels of Vitamin C than the seaweeds harvested from deeper waters. This may be due to the high antioxidant level needed for the seaweeds when exposed to the sun. According to a detailed review of the Vitamin C content of seaweeds by Cecilie et al., (2021) the Vitamin Egyptian J. of Phycol. Vol. 24, 2023 - 178 -

C content in seaweeds (dry wt.) is found to have a mean value of 0.773 mg g⁻¹ A study of taxonomical orders of species indicates that the green seaweeds of order Ulvales contain up to 3.0 mg g-1, whereas brown species of Fucales and Laminariales are found to have lower amounts of Vitamin C. During the present work, any prominent group-wise variation of Vitamin C content could not be observed. **Cecilie** *et al.* (**2021**) opine that Vitamin C content in seaweeds can vary due to biological, seasonal, locational, and treatment differences.

Iodine regulates the metabolism and proper growth of the human body and is an essential constituent of thyroid hormones that regulate major metabolic processes such as catabolism of carbohydrates, lipids and protein, cellular respiration, thermoregulation, intermediary metabolism, and nitrogen retention (**Abbaspour** *et al.*, **2014**; **Nunes** *et al.*, **2018**). Seaweeds are an excellent source of iodine; brown seaweeds contain the highest iodine content, with some species exceeding the RDI (150 μ g per day) (**Rajapakse & Kim, 2011**). Red and green seaweed species such as *Eucheuma cottoni*, *E. spinosum*, *Palmaria palmata*, *Porphyra* sp., and *Ulva lactuca* also contain iodine but at lower concentrations (**Zava & Zava, 2011; Nitschke & Stengel, 2016; Rasyid, 2017; Cherry** *et al.*, **2019**). Thus, the iodine requirement can be met by consuming seaweed or seaweed-based nutraceutical supplements (**Temjen, 2021**).

During the present study, the Iodine content was found to be high in almost all the algal species studied. It was found to vary from a minimum concentration (mg100g⁻¹ dry wt.) of 27.6 \pm 1.0 in *Ulva fasciata* to a maximum values of 86.9 \pm 1.0 and 86.8 \pm 1.0 in *Caulerpa peltata* and *Cladophora* sp. respectively (Chlorophyceae); from a minimum of 19.1 \pm 1.0 in *Sargassum tenerrimum* to a maximum of 36.0 \pm 1.0 in (Phaeophyceae), and from a minimum of 23.3 \pm 1.0 in *Cheilosporum spectabilis* to a maximum of 52.9 \pm 1.0 in *Acanthophora specifera and Champia parvula* (Rhodophyceae). The present values for iodine content in seaweeds are much lower than that reported by **Nida** Egyptian J. of Phycol. Vol. 24, 2023 - **179**-

et al. (2016) for those studied along the Pakistan coast. However, the present iodine values are comparable to the low levels (0.2 to 0.5 mg g⁻¹⁾ recorded in *Sargassum* spp. from the Gujarat coast by Ahmad *et al.* (1989). The present value of iodine content of 27.5 ± 1.0 mg $100g^{-1}$ is higher than that obtained for *Ulva Lactuca* (El-Tawil and Khalil, 1983). Global studies on iodine content in seaweeds from different coastal regions of the world indicate wide fluctuations in the values. Various factors like salinity, season, depth of the plants, water temperature, and post-harvest conditions are some of the factors influencing iodine content in seaweeds (Kravstova and Saenko, 1979). Figs. 8 to 10 furnish the vitamin C and iodine content in seaweed species belonging to different classes.

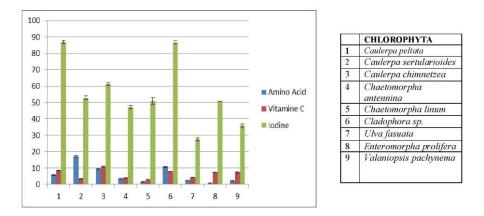


Fig. 8: Amino Acid, Vitamin C, and Iodine in green algae

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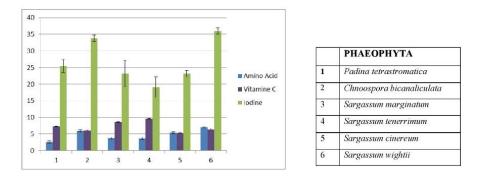


Fig. 9: Amino Acid, Vitamin C, and Iodine in brown algae

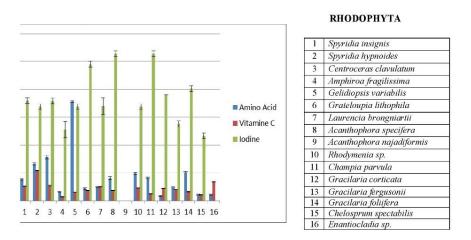


Fig. 10: Amino Acid, Vitamin C, and Iodine in red algae

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Conclusion

The overall observation of the present study shows that algae are rich sources of proteins, iodine, vitamins, and amino acids. These plants which are lavishly found in this ecosystem not only provide biomass by themselves but also have implications in the food and pharmaceutical industry as source materials in the preparation of food, food supplements, and fine chemicals. Nevertheless, only prolonged studies based on locations, depths, seasons, and various growth stages will help to derive any conclusive data on the nutritional composition of these edible marine plants. Besides not many studies have been undertaken on the toxic heavy metal contents of marine algae on Indian coasts, especially the southwest coast. Hence before incorporating these plants into the diet, it is essential to carry out an extensive study on their heavy metal profiling.

Acknowledgments

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References

- Abbaspour, N., Hurrell, R. and Kelishadi, R. (2014). Review on iron and its importance for human health. *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*, 19:164.
- Ahmad, J., Ganapathy, S.N., Siddiq T.O., and Hamdard, M.E. (1989). The distribution of elements in some plant species of the botanical kingdom. In: Elements in Health and Diseases. Said, M., Rahman, M.A., and Silva, L.A.D. eds. (Hamd. Univ. Press, Karachi), pp. 143 167.
- Angell, A.R., Mata, L., De Nys, R. and Paul, N.A. (2016). The protein content of seaweeds: a universal nitrogen-to-protein conversion factor of five. *Journal of Applied Phycology*, 28: 511–524.
- Arasaki, S. and Arasaki, T. (1983). Low Calorie, High Nutrition Vegetables from the Sea to Help You Look and Feel Better. (Vol. 60, Japan Publications, Tokyo), **pp.176**
- Aravindh S. and Ruban P. (2019). Seasonal variation of macroalgae in relation to physico-chemical parameters of coast of Kerala, India. *International Journal of Pharma and Bio Sciences*, 10 (3).
- Bligh, E.G., and Dyer W.J. (1959). A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*, 37: 911– 7.
- Boergesen, F. (1935). A list of marine algae from Bombay. Kgl. Danske Videnskabernes Selskab Biologiske Meddelesler, 12: 1-64
- Boergesen, F. (1938). Contribution to South Indian marine algal flora III. J. Indian Bot. Soc., 17: 205-242.

Egyptian J. of Phycol. Vol. 24, 2023

- 183 -

- Breitman, I., Saraf, N., Kakade, M., Yellumahanthi, K., White, M., Hackett, J.A. and Clements, R.H. (2011). The effects of an amino acid supplement on glucose homeostasis, inflammatory markers, and incretins after laparoscopic gastric bypass. *Journal of the American College of Surgeons*, 212: 617–625.
- Cecilie, W.N., Turid, R. and Susan, L.H. (2021). Vitamin C from Seaweed: A Review Assessing Seaweed as Contributor to Daily Intake. *Foods*, 19: 10(1):198.
- Černá, M. (2011). Seaweed proteins and amino acids as nutraceuticals. Advances in Food and Nutrition Research, 64: 297–312.
- Chakraborthy, S. and Santra, S.C. (2008). Biochemical composition of eight benthic algae collected from Sundarbans. *Indian Journal of Marine Science*, **37** (3): 329-332.
- Cherry, P., O'Hara, C., Magee, P.J., McSorley, E.M. and Allsopp, P.J. (2019). Risks and benefits of consuming edible seaweeds. *Nutrition Reviews*, 77: 307–329.
- Chennubhotla, V.S.K., Kaliaperumal, N., Kalimuthu, S& Ramalingam, J.R. (1991). Commercially important Seaweeds of India, their occurrence, chemical products, and use. Marine Fisheries Information Service, Technical and Extension Services, 107: pp 11-16.
- **Cordelia, F. (2021).** Goa's pilot seaweed farm explores the viability of this climate-smart algae. *https://india.mongabay.com/2021/10/goas-pilot-seaweed-farm-explores-viability-of-this-climate-smart-algae*

- 184 -

- Cotas, J., Marques, V., Afonso, M.B., Rodrigues, C.M. and Pereira, L. (2020). Antitumour potential of Gigartina pistillata carrageenans against colorectal cancer stem cell-enriched tumour spheres. Marine Drugs, 18: Article 50. DOI: 10.3390/md18010050.
- Daub, C.D., Mabate, B., Malgas, S. and Pletschke, B.I. (2020). Fucoidan from Ecklonia maxima is a powerful inhibitor of the diabetes-related enzyme, α-glucosidase. International Journal of Biological Macromolecules, 151: 412–420. DOI: 10.1016/j.ijbiomac.2020.02.161.
- Dawes, C. J. (1998). Physiological Ecology. In: Marine Botany, 2nd edition. (John Wiley and Sons, Inc., New York, USA), pp. 62-91.
- Dere, S., Dalkiran, N., Karacaoglu, D., Yildiz, G. and Dere, E. (2003). The determination of total protein, total soluble carbohydrate, and pigment contents of some macroalgae collected from Gemlik-Karacaali (Bursa) and Erdek-Ormanli (Balikesir) in the Sea of Marmara. *Turkey Oceanol.*, 45: 453–471.
- Desikachary, T.V. (1967). Seaweed resources of India. In: V. Krishnamurthy (ed.) Proceedings of the seminar on sea, salt, and plants, CSMCRI, Bhavnagar, India Pp. 7-24
- Dhargalkar, V.K., Jagtap, T.G. and Untawale, A.G. (1980). Biochemical constituents of seaweeds along the Maharashtra coast. *Indian J. of Marine Science*, 9: 297-299.
- Dinesh, G., Sekar M. and Kannan, R. (2007). Nutritive properties of seaweeds of Gulf of Mannar, Tamil Nadu. *Seaweed Res. Utilization*, 29: 125-132.
- El-Tawil, B.A.H. and Khalil, A.N. (1983). Chemical constituents of some algal species from Abu-Qir Bay, *Egypt. J. Facul. Mar. Sci.* 3: 1404H: 85 94.

- 185 -

- Fleurence, J., Chenard, E. and Lucon, M. (1999). Determination of the nutritional value of proteins obtained from Ulva *armoricana*. *Journal of Applied Phycology*, 11: 231–239.
- **Fujiwara-Arasaki, T., Mino, N. and Kuroda, M**. (**1984**). The protein value in human nutrition of edible marine algae in Japan. *Hydrobiologia*, 34: 27-50.
- Ganesan, A. R., Tiwari, U., and Rajauria, G. (2019). Seaweed nutraceuticals and their therapeutic role in disease prevention. *Food Sci. Hum. Wellness*, 8:252–263.doi: 10.1016/j.fshw.2019.08.001
- Holdt, S.L. and Kraan, S. (2011). Bioactive Compounds in Seaweed: Functional Food Applications and Legislation. *Journal of Applied Phycology*, 23: 543-597.
- Hayashi, Y. and Seino, Y. (2018). Regulation of amino acid metabolism and α- cell proliferation by glucagon. *Journal of Diabetes Investigation*, 9: 464–472.
- Hyman Rosen. (1957). A modified ninhydrin colorimetric analysis for amino acids. Archives of Biochemistry and Biophysics, 67 (1): 10-15.
- Iyengar, M.O.P. (1927). Krusadi island Flora. Bull. Madras. Gov. Mus. News, 1:85-188
- Kaliaperumal, N. (2006). Method for assessment of seaweed Resources and identification of marine algae. National training workshop on marine and coastal biodiversity assessment for conservation and sustainable utilization. Suganthi Devadason Marine Research Institute Special Research Publication, No. 10: 72-82

Egyptian J. of Phycol. Vol. 24, 2023 - 186 -

- Kawai, M., Sekine-Hayakawa, Y., Okiyama, A. and Ninomiya, Y. (2012). Gustatory sensation of L- and d- amino acids in humans. *Amino acids*, 43(6): 2349- 2358.
- Kravtsova, Y. and Saenko, G.N. (1979). Biological aspects of iodine behaviour during interaction of Algae with seawater. E.V. K. ed. Vaimodeistvie Vodoi Zhivym Veshchestvom Tr. Mezhdunar. Simp. 1975, Publ. 1, Moscow, pp. 146–152
- Krishnamurthy, V. (1971). Seaweed resources of India and their utilization. Seaweed Research Utilization, 1: 55-67.
- Lowry, O.H., Rosenbrough, N.J., Farr, A.L. and Randall, R.J. (1951). Protein measurement with the Folin phenol reagent. J. Biol. Chem., 193:265-275.
- Machado, M., Machado, S., Pimentel, F. B., Freitas, V., Alves, R.C. and Oliveira, M.B.P.P. (2020). Amino acid profile and protein quality assessment of macroalgae produced in an integrated multi-trophic aquaculture system. *Foods*, 9(10): 1382.
- Mahadevan, A. and Sridhar, R. (1996). Methods in physiological plant pathology, Sivakami Publication, 4th edition, Madras, **pp.316**.
- Manivannan, K., Thirumaran, G., Devi, G.K., Hemalatha, A. and Anantharaman, P. (2008a). Biochemical composition of seaweeds from Mandapam Coastal regions along Southeast Coast of India. *Am-Euras. J. Bot.*, 1:32-37.
- Manivannan, K., Devi, G.K., Thirumaran, G., Anantharaman, P. (2008b). Mineral composition of marine macroalgae from Mandapam Coastal regions; Southeast Coast of India. *Am-Euras J. Bot.* 1: 58-67.

Egyptian J. of Phycol. Vol. 24, 2023 - 187 -

- Marinho-Soriano, E., Fonseca, P.C., Carneiro, M.A.A. and Moreira, W.S.C. (2006). Seasonal Variation in the Chemical Composition of Two Tropical Seaweeds. *Bioresource Technology*, 97: 2402-2406.
- Martin, D.F. (1969). Marine chemistry; Analytical methods; Marcel Decker, INC New York, Pp: 280.
- Meghanath, P., Alexander, C., Ruth, G., Meital, K., Omir, N., Michael, G., Alvaro, I., Yoav, D. Livney and Alexander G. (2019). Starch from the sea: The green macroalga *Ulva Ohnoi* as a potential source for sustainable production in the marine biorefinery. *Algal Research*, 37: 215 – 227.
- Maya, S. (1990). Biocoenosis of intertidal Rock Pools with special reference macrophytes. PhD thesis submitted to the University of Kerala, India, pp. 316
- Maya Subramoni. (2010). Algal resources of Southwest coast of India Systematic study and evaluation as food and food supplements. Report submitted to Department of Science and Technology, Gov. of India, pp. 32
- Mellouk Z., Benammar I., Krouf D., Goudjil M., Okbi M. & Malaisse W. (2017). Antioxidant properties of the red alga *Asparagopsis taxiformis* collected on the North West Algerian coast. *Experimental and Therapeutic Medicine*, 13: 3281–3290. DOI: 10.3892/etm.2017.4413
- Misurcova, L. (2012). Chemical composition of seaweeds part II: isolation and chemical properties of Molecules Derived from Seaweeds. Handbook of Marine Macroalgae: Biotechnology and Applied Phycology, 1st Edition. Se-Kwon Kim ed. (John Wiley & Sons, Ltd.), pp. 173-192

- 188 -

- Nair, N.B., Sobha, V., Rathi Ammal, M., Chandran, R., Maya, S., Suryanarayan, H. (1986). Algal resources of Kerala coast II – An updated list of Indian marine algae. *Aquatic Biology*, 6: 25-52
- Nair, N.B., Sobha, V., Rathi Ammal, M., Chandran, R., Maya, S., Suryanarayan, H. and Vasudevan Nair, T. (1991). Algal resources of Kerala coast V- Protein and lipid contents of some marine algae. Proceedings of the Kerala Science Congress, Calicut, pp.103-105
- Necas, J. and Bartosikova, L. (2013). Carrageenan: a review. Veterinarni Medicina, 58: 187–205. DOI: 10.17221/6758-VETMED.
- Nehal, A. Osman, Islam, M. El- Manwy and Abeer Sh. Amin. (2011). Nutritional composition and mineral content of five macroalgae from Red Sea. *Egyptian J. Phycology*, **12-89-102**.
- Nida, A. K., Aisha, K., and Ghazala, S. (2016). Extraction and estimation of iodine from brown seaweeds, of Karachi coast, Pakistan. *Pakistan Journal of Marine Sciences*, 25(1&2): 15-25.
- Nitschke, U. and Stengel, D.B. (2016). Quantification of iodine loss in edible Irish seaweeds during processing. *Journal of Applied Phycology*, 28: 3527–3533. DOI: 10.1007/s10811-016-0868-6.
- Nunes, N., Rosa, G.P., Ferraz, S., Barreto, M.C., and de Carvalho, M.P. (2019). Fatty acid composition, TLC screening, ATR–FTIR analysis, anti-cholinesterase activity, and in vitro cytotoxicity to A549 tumour cell line of extracts of 3 macroalgae collected in Madeira. *Journal of Applied Phycology*, 1–13.
- O'Connor, J., Meaney S., Williams, G.A., and Hayes, M. (2020). Extraction of Protein from Four Different Seaweeds Using Three Different Physical Pre-Treatment Strategies. *Molecules*, 25(8): 2005.

- 189 -

- Ortiz, Romero, N., Robert, P., Araya, J., Lopez'-Hernandes, J., Bozzo, C., et.al. (2006). Dietary fibre, amino acid fatty acid, and tocopherol contents of the edible seaweeds Ulva lactuca and Durvillaea Antarctica, *Food Chemistry*, 99(1): 98-104.
- Paul M. A., Christopher, I.R. Gill, Mariel Brooks, Ross Campbell and Ian, R. Rowland. (2007). Nutritional Value of Edible Seaweeds. *Nutrition Reviews*, 65(1): 535-543
- Penalver, R. M., Jose M. L., Gaspar, R.s, Ryszard A. (2020). Seaweeds as Functional Ingredient for a Healthy Diet. *Marine Drugs*, 18(6): 301
- Pereira, L. (2011). A review of the nutrient composition of selected edible seaweeds. Seaweed: Ecology, Nutrient Composition and Medicinal Uses: 15–47.
- Pereira, L. (2018a). Seaweeds a source of bioactive substances and skin care therapy cosmeceuticals, algotheraphy, and thalassotherapy. *Cosmetics*, 5: 68.
- Pereira, L. (2018b). Therapeutic and nutritional uses of algae. CRC Press, New York, New York, USA. 672 pp.
- Rajapakse, N., & Kim S.K., (2011). Nutritional and digestive health benefits of seaweed. In *Advances in Food and Nutrition Research*, 64: 17–28.
- Rameshkumar, S., Ramakritinan, CM., Yokeshbabu, M. (2012). Proximate composition of some selected seaweeds from Palk Bay and Gulf of Mannar, Tamil Nādu, India. Asian Journal of Biomedical and Pharmaceutical Sciences, 3(16): 1-5

Egyptian J. of Phycol. Vol. 24, 2023 - 190 -

- Reddy, C.R.K., Subbarao, P.V., Ganesan, M., Eswaran, S., Zaidi, H., Mantri,
 V.A. (2014). The Seaweed resources of India. In: Critchley, A.T., Ohno,
 M. and Largo D.B. (eds.). World Seaweed Resources, ETI Information
 Services Ltd., Wokingham, Berkshire, U.K.
- Rasyid N.Q. (2017). Analysis of iodine content in seaweed and estimation of iodine intake. *Marina Chimica Acta*, 18: 26–30.
- Rosenberg, C. and Ramus, J. (1982). Ecological growth strategies in the seaweeds *Gracilaria foliifera* (Rhodophyceae) and *Ulva* sp. (Chlorophyceae): soluble nitrogen and reserve carbohydrates. *Marine Ecology-Progress Series*, 8: 233 241.
- Sarker, S., Siddique, M.A.B., Bithi, U.H., Rahman, M.M., Rahman, M.S., Akter, M. (2021). Diseases, metals, and bioactive compounds in seaweeds of Bangladesh. *Reg. Stud. Mar. Sci.*, 48 :102021.
- Shannon, E., Abu-Ghannam, N. (2019). Seaweeds as Neutraceuticals for Health and Nutrition. *Phycologia*, 58: 563-577
- Shynu, S., Prabha, D. L., and Thomas G. (2012). Ecology of seaweeds along Thirumullavaram shoreline, Kerala. J. Recent Trends Biosci., 2(2): 20-25.
- Sitakara Rao, V. and Tipnis, U.K. (1967). Chemical constituents of marine algae from Gujarat coast. Proceedings of Seminar on Sea Salt and Plants, CSMCRI, Bhavnagar, pp. 277-288
- Skrovankova, S. (2011). Seaweed vitamins as nutraceuticals. In Advances in Food and Nutrition Research; Elsevier: Amsterdam, The Netherlands, 64: 357–369.
- Strickland, J.D.H. and T.B.A. Parsons, (1972). Practical handbook of seawater analysis. J. Fish. Res. Bd. Canada.Bull.,167: 311.

- 191 -

- Sushant, V. R. and Madaiah, R. (2014). Seasonal Assessment of Hydrographic variation and Phytoplanktonic community in the Arabian Sea waters of Kerala, Southwest Coast of India. *Brazilian Journal of Oceanography*, 62(4): 279-293.
- Taboada, M., Millán, R. and Miguez, M., (2013). Nutritional value of the marine algae wakame (Undaria pinnatifida) and nori (Porphyra purpurea) as food supplements. Journal of Applied Phycology, 25: 1271– 1276
- Tanna, B., Mishra, A. (2019). Nutraceutical Potential of Seaweed Polysaccharides: Structure, Bioactivity, safety, and toxicity. *Compr. Rev. Food Sci. Food Saf.*, 18(3): 817-831
- Tee, E.S, S.I Young and Ho S.K. and SitiMizura, S. (1988). Determination of vitamin C in fresh fruits and vegetables using dye titration and microfluorometrics. *Pertanike*, 11(1): 39-44
- Temjen I. (2021). Nutritional value of marine seaweeds and their potential to serve as nutraceutical supplements. Phycologia, 60(6): 534 546
- Tseng, C.K. (2004). Past, present, and future of phycology in China. *Hydrobiologia*, 512:11–20
- **UNICEF- MOH**. (1995). The miracle of iodized salt: Ethiopia's commitment of USI, circular, No.9.
- Untawale, A.G., Agadi, V.V., Dhargalkar, V.K. (1983). List of Marine Algae from India. National Institute of Oceanography, Goa.
- Wang W., Wang S.-X. & Guan H.-S. (2012). The antiviral activities and mechanisms of marine polysaccharides: an overview. *Marine Drugs* 10: 2795–2816. DOI: 10.3390/md10122795.

Egyptian J. of Phycol. Vol. 24, 2023 - 192 -

- Wee, M.S. and Henry, C.J. (2020). Reducing the glycemic impact of carbohydrates on foods and meals: strategies for the food industry and consumers with special focus on Asia. *Comprehensive Reviews in Food Science and Food Safety*, 19: 670–702. DOI: 10.1111/1541-4337.12525
- Yemm, W., and Willis, A.J. (1954). The Estimation of Carbohydrates in Plant Extracts by Anthrone. *Biochemical Journal*, 57: 508-514.
- Yemm, E.W., and Cocking, E.C. (1955). The Determination of Amino Acids with Ninhydrin. *Analyst*, 80: 209-213.
- Zava, T.T. and Zava, D.T. (2011). Assessment of Japanese iodine intake based on seaweed consumption in Japan: a literature-based analysis. *Thyroid Research*, 4: Article 14. DOI: 10.1186/1756-6614-4-14
- Zhang, L., Liao, W., Huang, Y., Wen, Y., Chu, Y., Zhao, C. (2022). Global seaweed farming and processing in the past 20 years. *Food Prod. Process. Nutr.*, 4: 23.

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